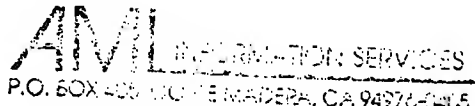


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**(54) Title:** RECEPTOR FOR ONCOSTATIN M AND LEUKEMIA INHIBITORY FACTOR

**(57) Abstract**

A receptor protein comprising a gp130 polypeptide linked to a single-chain leukemia inhibitory factor receptor (LIF-R) polypeptide is capable of binding both oncostatin M and leukemia inhibitory factor (LIF). The receptor protein binds LIF with greater affinity than does the single-chain LIF-R polypeptide alone. The receptor may be produced as a fusion protein in recombinant cells. The gp130 polypeptide binds oncostatin M, but with lower affinity than does the inventive receptor protein.

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**TITLE**

Receptor for Oncostatin M and Leukemia Inhibitory Factor

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**BACKGROUND OF THE INVENTION**

Receptors that bind specific molecules (e.g., a hormone, drug, cytokine, or biochemical) have been identified on a multitude of cell types. Receptors are found on the cell surface or, in the case of soluble receptors, are released into the serum. Effort has been directed toward isolation and characterization of a number of receptors in order to study their physiological roles and to explore possible therapeutic uses. The binding of a particular target molecule by a soluble receptor administered to a patient may alleviate disorders mediated by the target molecule.

Certain receptors have been found to comprise two separate polypeptide chains associated in the form of a complex. Such two-chain receptors often bind the target molecule with greater affinity than that exhibited by one of the chains alone.

Leukemia inhibitory factor (LIF) is a polypeptide hormone that plays a central role in the regulation of diverse adult and embryonic systems. LIF acts on a variety of cell types and has multiple biological activities. The diversity in biological activity is reflected in the various synonyms of LIF, which include hepatocyte stimulating factor III (Baumann and Wong, *J. Immunol.* 143:1163 [1989]); cholinergic nerve differentiation factor (Yamamori et al., *Science* 246: 1412 [1990]); melanoma-derived lipoprotein lipase inhibitor (Mori et al., *Biochem. Biophys. Res. Comm.* 160:1085 [1989]); human interleukin for DA cells (Moreau et al., *Nature* 336:690 [1988]); differentiation factor (Tomida et al., *J. Biol. Chem.* 259:10978 [1984]); differentiation inhibitory factor (Abe et al., *J. Biol. Chem.* 264: 8941 [1989]); differentiation inhibitory activity (Smith and Hooper, *Devel. Biol.*, 121:1 [1987]); and differentiation retarding factor (Koopman and Cotton, *Exp. Cell. Res.* 154:233 [1984]).

The cloning of a leukemia inhibitory factor receptor (LIF-R) has been reported by Gearing et al. in *EMBO J.* 10:2839 (1991). This recombinant single-chain LIF-R polypeptide binds LIF, but with lower affinity than the naturally occurring LIF

receptors found on certain normal cells. A receptor that binds LIF with higher affinity than that exhibited by this cloned single chain LIF-R would be desirable for certain applications.

5 Oncostatin M is a secreted single-chain polypeptide cytokine that regulates the growth of certain tumor-derived and normal cell lines. Oncostatin M is produced by activated lymphoid cells. A number of cell types have been found to bind the oncostatin M protein. See, for example, Linsley et al., *J. Biol. Chem.*, 264: 4282 (1989). However, the isolation and characterization of an oncostatin M receptor have not been reported.

10

### SUMMARY OF THE INVENTION

The present invention provides a receptor that has the property of binding both oncostatin M and leukemia inhibitory factor (LIF). The receptor comprises gp130 linked (preferably covalently) to leukemia inhibitory factor receptor (LIF-R). The gp130 polypeptide may be covalently linked to the LIF-R polypeptide by any suitable means, such as *via* a cross-linking reagent or a polypeptide linker. In one embodiment of the invention, the receptor is a fusion protein produced by recombinant DNA technology. Disorders mediated by either oncostatin M or LIF may be treated by administering a therapeutically effective amount of the inventive receptor to a patient afflicted with such a disorder.

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### BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a graph presenting the results of an LIF binding assay. Host cells transfected with vector(s) encoding gp130 or LIF-R were assayed for the ability to bind LIF, as described in example 1.

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Figure 2 is a graph presenting the results of an oncostatin M binding assay. Host cells transfected with vector(s) encoding gp130 or LIF-R were assayed for the ability to bind oncostatin M, as described in example 2.

Figure 3 is a graph depicting low affinity binding of oncostatin M to host cells transfected with a gp130 encoding expression vector, as described in example 2.

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Figure 4 schematically depicts a receptor of the present invention wherein Fc polypeptides derived from an antibody are used to link a gp130 fragment to an LIF-R fragment.

Figure 5 presents composite DNA and encoded amino acid sequences of a full length LIF-R, determined by comparing the sequences of cDNA and genomic clones. The signal peptidase cleavage site is marked with a vertical arrow. The transmembrane region is heavily underlined. Potential N-linked glycosylation sites are marked with

35

asterisks. Hallmark residues associated with the hematopoietin family of receptors are shown boxed. The horizontal arrow marks the point at which genomic sequence was used to derive the 3' coding region of LIF-R, since the cDNA clones employed in determining this sequence terminated with a stretch of A nucleotides at this point.

5        Figure 6 presents the DNA and deduced amino acid sequences of cloned gp130 cDNA as reported by Hibi et al. in *Cell* 63:1149 (1990). A predicted signal sequence is underlined. The thick underline indicates a presumed transmembrane region. The sets of asterisks identify possible N-glycosylation sites.

10        Figure 7 presents Scatchard analyses that demonstrate the interaction of a soluble gp130/Fc fusion protein with soluble LIF-R/Fc in binding LIF and oncostatin M, as described in example 7.

### **DETAILED DESCRIPTION OF THE INVENTION**

15        The present invention provides a receptor comprising gp130 covalently linked to leukemia inhibitory factor receptor (LIF-R). In another embodiment of the invention, the receptor comprises gp130 non-covalently complexed with LIF-R. The receptor is capable of binding oncostatin M, and also binds leukemia inhibitory factor (LIF). The receptor thus is useful for treating disorders mediated by either oncostatin  
20        M or LIF.

      The gp130 may be covalently linked to the LIF-R by any suitable means, such as *via* a cross-linking reagent or a polypeptide linker. The gp130 and LIF-R proteins are covalently linked in a manner that does not interfere with the resulting receptor's ability to bind oncostatin M and LIF. In one embodiment of the invention, the receptor  
25        is a fusion protein produced by recombinant DNA technology.

      Non-covalent bonding of gp130 to LIF-R may be achieved by any suitable means that does not interfere with the receptor's ability to bind oncostatin M and LIF. In one approach, a first compound is attached to LIF-R and a second compound that will non-covalently bond to the first compound is attached to gp130. Examples of  
30        such compounds are biotin and avidin. The receptor is thus formed through the non-covalent interactions of biotin with avidin. In one embodiment of the invention, LIF-R and gp130 are recombinant polypeptides, each purified from recombinant cells and then non-covalently bonded together to form the receptor. A host cell may be transformed with two different expression vectors such that both LIF-R and gp130 are produced by  
35        the recombinant host cell. LIF-R and gp130 (one or both of which are soluble fragments as described below) produced by such transformed host cells may associate to form a complex through non-covalent interactions.

"Leukemia inhibitory factor receptor" (LIF-R) refers to a protein (a cytokine receptor) that is present on the surface of various hematopoietic cells, including monocyte-macrophages and megakaryocytes, and on non-hematopoietic cells, including osteoblasts, placental trophoblasts, and liver parenchymal cells. LIF-R is capable of binding leukemia inhibitory factor (LIF) molecules and plays a role in transducing the signal provided by LIF to a cell. In the absence of any species designation, LIF-R refers generically to mammalian LIF-R, which includes, but is not limited to, human, murine, and bovine LIF-R.

The cloning of human and murine leukemia inhibitory factor receptors (LIF-R), each a single polypeptide chain, has been reported by Gearing et al. in *EMBO J.* 10:2839 (1991), which is hereby incorporated by reference in its entirety. The DNA sequence of a human LIF-R cDNA clone and the amino acid sequence encoded thereby are shown in SEQ ID NO: 5 and SEQ ID NO: 6. This cloned human cDNA encodes an N-terminal fragment of human LIF-R that includes (in order from N-terminus to C-terminus) a 44-amino acid signal sequence (amino acids -44 to -1), the entire extracellular region, a transmembrane region (the first amino acid of which is amino acid number 790 of SEQ ID NO: 5) and a portion of the cytoplasmic domain. The C-terminus of the fragment includes amino acids encoded by a poly-A segment and by a linker employed in vector construction, as described in Gearing et al., *supra*. The term "transmembrane region" as used herein refers to a string of hydrophobic amino acids positioned between the extracellular domain and the cytoplasmic domain of the protein. A plasmid vector containing the above-described cloned human LIF-R cDNA is designated pHLIFR-65 and has been deposited in *E. coli* host cells with the American Type Culture Collection on December 11, 1990 (ATCC accession no. 68491). The DNA and amino acid sequences of a full length native human LIF-R (determined by comparing the sequences of cDNA and genomic clones) have been reported by Gearing et al. *supra* and are presented herein in Figure 5.

The LIF-R encoded by the cloned cDNA (SEQ ID NO: 6) contains the entire extracellular region of LIF-R (the domain believed to be responsible for the LIF-binding activity), and binds LIF, but with lower affinity than does a naturally occurring LIF receptor found on certain normal cells. Additionally, oncostatin M competes with LIF for binding to naturally occurring high affinity LIF receptors on certain cell types (Gearing et al., *New Biologist*, 4:61, 1992) but did not bind to the above-described cloned LIF-R expressed in COS cells.

In order to investigate the possible existence of a high affinity converting subunit for the cloned single polypeptide chain LIF-R, host cells were co-transfected with the LIF-R encoding plasmid pHLIFR-65 and with pools from a human placental

cDNA library (also contained in an expression vector). The co-transfected cells were assayed for the ability to bind radiolabeled oncostatin M.

A positive cDNA pool was subdivided and the procedure repeated to isolate a single cDNA clone designated B10G that conferred the ability to bind oncostatin M on  
5 cells co-transfected with B10G and the LIF-R encoding plasmid pHLIFR-65. The co-transfected cells also were found to bind LIF with higher affinity than cells transfected with pHLIFR-65 alone. Host cells transfected with B10G alone exhibited low affinity oncostatin M binding sites. The B10G cloned cDNA was sequenced and found to encode a protein that is known as gp130.

10 Thus, it has now been found that a receptor comprising both LIF-R and gp130 binds LIF with higher affinity than does the single-chain LIF-R polypeptide alone. The improved LIF binding of LIF-R in combination with gp130 is described in example 1 below and depicted in Figure 1.

Although LIF does not bind to either high- or low-affinity oncostatin M  
15 receptors, it has now been found that oncostatin M binds to the receptors of the present invention comprising LIF-R and gp130. Oncostatin M binding is described in example 2 below and depicted in Figure 2.

A protein known as gp130 has been purified from cellular sources that include placental tissue and a myeloma cell line U266. A number of additional cell types have  
20 been found to express gp130 mRNA, as reported by Hibi et al., in *Cell* 63:1149 (1990). gp130 has been reported to be involved in the formation of high affinity interleukin-6 binding sites and in IL-6 signal transduction (Hibi et al. *supra*). The cloning and expression of cDNA encoding a full length gp130 protein has been reported by Hibi et al., *supra*, which is hereby incorporated by reference in its entirety.  
25 The DNA and deduced amino acid sequences reported by Hibi et al. for the gp130 cloned cDNA are presented herein in figure 6. The gp130 amino acid sequence may vary from that reported by Hibi et al., e.g., leucine may be substituted for valine at position 8 in the signal sequence (numbering is as shown in Figure 6). This amino acid substitution may be attributable to genetic polymorphism (allelic variation among  
30 individuals producing the protein), and results from the presence of C rather than G at nucleotide position 22.

As used herein, the term LIF-R includes variants and truncated forms of native LIF-R proteins that possess the desired LIF-binding or signal transducing activity. Likewise, the term gp130 as used herein includes variants and truncated forms of the  
35 native gp130 protein that retain the desired biological activity. For gp130, the desired biological activity includes binding of oncostatin M; conferring on the inventive receptor the ability to bind oncostatin M; and increasing the affinity of the inventive

receptor for LIF, compared to the LIF binding affinity of the single-chain LIF-R polypeptide alone. Specifically included are truncated, soluble or fusion forms of LIF-R and gp130, as described below. Variants produced by adding, substituting, or deleting amino acid(s) in the native sequence are discussed in more detail below.

5 One example of an LIF-R polypeptide that may be employed is that encoded by the cDNA clone designated pHLIF-R-65 (SEQ ID NO: 5), as described by Gearing et al., *supra* and in example 3 below. Alternatively, a fragment comprising amino acids 1 to 945 of SEQ ID NO:5 may be employed. Amino acid 945 is the last LIF-R-specific amino acid of the polypeptide encoded by clone pHLIF-R-65, before the poly-A  
10 nucleotide segment believed to result from oligo(dT) priming at an internal site in the mRNA during preparation of the hLIF-R cDNA. (See Gearing et al., *EMBO J.*, *supra*. at page 2840, column one.)

Other examples of LIF-R polypeptides that may be employed in the inventive  
15 receptors include those lacking all or part of the transmembrane region or the cytoplasmic domain of the protein. Suitable LIF-R polypeptides thus include those containing amino acids 1-x or, when the signal sequence is not desired, amino acids 45-x of the full length LIF-R sequence depicted in Figure 5, wherein x represents an integer from 833 to 1096. Amino acid number 833 is the last amino acid of the extracellular domain (i.e., before the start of the transmembrane region.) Polypeptides  
20 terminating in amino acid number 1096 lack the last C-terminal amino acid of the full length protein. The desirability of including the signal sequence depends on such factors as the position of LIF-R in a fusion protein, as discussed below, and the intended host cells when the receptor is to be produced *via* recombinant DNA technology. Note that the numbering of amino acids in Figure 5 (taken from Gearing et  
25 al., *supra*) differs from that of SEQ ID NO: 5 because the first amino acid of the signal sequence is designated amino acid number 1 in Figure 5 but is designated -44 in SEQ ID NO: 5. Other polypeptides may be chosen with regard to sequences that are conserved in the hematopoietin receptor family, (i.e., chosen to include the boxed sequence(s) shown in Figure 5.)

30 One example of a suitable gp130 polypeptide is that encoded by cDNA cloned into plasmid vector pDC303 to produce a plasmid designated B10G. The source of mRNA used in producing the cDNA was human placental tissue. Plasmid B10G in *E. coli* strain DH5 $\alpha$  host cells was deposited with the American Type Culture Collection, Rockville, Maryland, on November 14, 1991, and assigned ATCC accession number  
35 68827.

The DNA sequence of the gp130 cDNA contained in plasmid B10G and the amino acid sequence of the gp130 protein encoded by the cloned cDNA are presented in

SEQ ID NO: 1 and SEQ ID NO: 2. The protein comprises (in order from the N-terminus to the C-terminus) a 22-amino acid signal sequence, complete extracellular domain (amino acids 1-597), a transmembrane region (beginning with amino acid 598), and a partial cytoplasmic domain (amino acids 621-686). This truncated gp130 polypeptide differs from the equivalent portion of the Hibi et al. protein in that the eighth amino acid of the signal sequence is leucine rather than valine, as discussed above.

Another example of a suitable gp130 polypeptide comprises amino acids 1 to 496 of the SEQ ID NO: 1, which includes all of the cysteine residues found in the extracellular domain of the protein, and also contains a complete fibronectin domain. Additional examples of gp130 polypeptides are those comprising amino acids 1-298 or 98-298 of SEQ ID NO: 1.

Other gp130 polypeptides lacking all or part of the transmembrane region and/or cytoplasmic domain may be employed. Suitable gp130 polypeptides thus include those containing amino acids 1-x or, when the signal sequence is not desired, amino acids 23-x of the Figure 6 sequence, wherein x represents an integer from 619 to 917. The first amino acid of the transmembrane region is the alanine residue at position 620 in Figure 6. Polypeptides terminating at amino acid 917 lack the last C-terminal amino acid of the full length protein presented in Figure 6. Note that the numbering of amino acids in Figure 6 (taken from Hibi et al., *supra*) differs from that shown in SEQ ID NO: 1 and NO:2 because the first amino acid of the signal sequence is designated amino acid number 1 in Figure 6 but is designated -22 in SEQ ID NO: 1. Regions of the gp130 protein corresponding to domains that are conserved among certain receptors are discussed by Hibi et al, *supra*, at page 1150, column 2, and page 1151, column 1. Other truncated gp130 polypeptides chosen to include these conserved regions may be employed.

Preferred LIF-R and gp130 polypeptides are those which are soluble. In one embodiment of the present invention, the receptor comprises soluble LIF-R covalently attached to soluble gp130. "Soluble LIF-R" as used in the context of the present invention refers to polypeptides that are substantially similar in amino acid sequence to all or part of the extracellular region of a native LIF-R and that, due to the lack of a transmembrane region that would cause retention of the polypeptide on a cell membrane, are secreted upon expression. The soluble LIF-R polypeptides that may be employed retain the ability to bind LIF or, by competitively binding LIF, inhibit LIF signal transduction activity via cell surface bound LIF-R proteins. Soluble LIF-R may also include part of the transmembrane region or part of the cytoplasmic domain or other sequences, provided that the soluble LIF-R protein is capable of being secreted.

Likewise, the term "soluble gp130" as used herein refers to proteins that are substantially similar in amino acid sequence to all or part of the extracellular region of a native gp130 and are secreted upon expression but retain the desired biological activity. Soluble gp130 may include part of the transmembrane region, cytoplasmic domain, or  
5 other sequences, as long as the polypeptide is secreted.

Soluble LIF-R and soluble gp130 may be identified (and distinguished from their non-soluble membrane-bound counterparts) by separating intact cells which express the desired protein from the culture medium, e.g., by centrifugation, and assaying the medium (supernatant) for the presence of the desired protein. The culture  
10 medium may be assayed using procedures which are similar or identical to those described in the examples below. The presence of LIF-R or gp130 in the medium indicates that the protein was secreted from the cells and thus is a soluble form of the desired protein. Soluble LIF-R and soluble gp130 may be naturally-occurring forms of these proteins. Cloning of a naturally-occurring soluble murine LIF-R is reported in  
15 Gearing et al., *supra*. Alternatively, soluble fragments of LIF-R and gp130 proteins may be produced by recombinant DNA technology or otherwise isolated, as described below.

The use of soluble forms of LIF-R and gp130 is advantageous for certain applications. Purification of the proteins from recombinant host cells is facilitated,  
20 since the soluble proteins are secreted from the cells. Further, soluble proteins are generally more suitable for intravenous administration and may exert their therapeutic effect (binding LIF and oncostatin M) in the bloodstream.

Soluble LIF-R polypeptides include those comprising the signal sequence and entire extracellular domain (amino acids -44 to 789 of SEQ ID NO: 5) or lacking the  
25 signal sequence but containing the entire extracellular domain (amino acids 1 to 789 of SEQ ID NO: 5). Soluble gp130 polypeptides include those comprising the signal sequence and entire extracellular domain (amino acids -22 to 597 of SEQ ID NO: 1) or lacking the signal sequence but containing the entire extracellular domain (amino acids 1 to 597 of SEQ ID NO: 1). The preparation and use of these soluble polypeptides in  
30 receptors of the present invention is described in examples 3-5.

Other soluble LIF-Rs are truncated upstream of the transmembrane region, but preferably include that portion of the protein that contains the residues conserved among the members of the hematopoietin receptor family (shown boxed in Figure 5), i.e., amino acids 11-479 of SEQ ID NO:6. The N-terminus of such soluble LIF-Rs is  
35 any of amino acids 1-11 (or -44 when the native signal sequence is included), and the protein extends to a C-terminus selected from any of amino acids 479 through 789. Two such soluble proteins comprise amino acids -44 - 702, 1 - 702, -44 - 775, or 1 -

755 of SEQ ID NO:6. Constructs encoding these proteins may be prepared by techniques that involve cleaving the human LIF-R cDNA of the above-described clone pHLIFR-65 (ATCC 68491) with the restriction endonucleases *Asp718* and *Xmn1* or with *Asp718* and *Bsp1286I*. *Asp 718* cleaves the vector upstream of the inserted LIF-R-encoding cDNA. *Xmn1* cleaves within the codon for Asp at position 702 (generating blunt ends) and *Bsp1286I* cleaves just 3' of the codon for Val at position 775 of SEQ ID NO:5. If desired, an oligonucleotide may be ligated to the 3' end of the *Asp718/Bsp1286I* fragment to extend the LIF-R sequence, e.g., through amino acid number 789. An oligonucleotide also may be ligated to the 3' end of a LIF-R fragment to add the first two amino acids of the Fc polypeptide described in example 5, and a Bgl II site useful for attaching the rest of the Fc sequence downstream of the LIF-R sequence.

Additional soluble human LIF-Rs comprise amino acids 1-678 or 1-680 of SEQ ID NO:6. When the human and murine LIF-R amino acid sequences disclosed in Gearing et al., *EMBO J.*, *supra*, are aligned (with gaps introduced to maximize identity between the two sequences), amino acid 680 of the human sequence is aligned with the last amino acid of the murine protein, and amino acid 678 is the last amino acid of the human sequence that is identical to a corresponding amino acid in the murine sequence. Since the murine protein binds LIF, the murine LIF-R contains that portion of the protein required for LIF binding.

An additional example of a soluble gp130 polypeptide comprises amino acids -22 to 582 of SEQ ID NO:2. An expression vector encoding such a protein was constructed in example 7. Soluble LIF-R and gp130 polypeptides also include those from which fibronectin type III (FNIII) domains have been deleted. From one to all of the FNIII domains may be deleted, providing the advantage of reducing the size of the protein. Preparation of such LIF-R and gp130 proteins is described in example 8.

Truncated LIF-R and gp130, including soluble polypeptides, may be prepared by any of a number of conventional techniques. In the case of recombinant proteins, a DNA fragment encoding a desired fragment may be subcloned into an expression vector. Alternatively, a desired DNA sequence may be chemically synthesized using known techniques. DNA fragments also may be produced by restriction endonuclease digestion of a full length cloned DNA sequence, and isolated by electrophoresis on agarose gels. Linkers containing restriction endonuclease cleavage site(s) may be employed to insert the desired DNA fragment into an expression vector, or the fragment may be digested at cleavage sites naturally present therein. Alternatively, proteins may be fragmented using proteolytic enzymes, for example, and the desired truncated polypeptide isolated from the digestion mixture using reversed phase HPLC.

The well known polymerase chain reaction procedure also may be employed to isolate a DNA sequence encoding a desired protein fragment. This technique is illustrated in examples 3-5 below.

5 In another approach, enzymatic treatment (e.g., using Bal 31 exonuclease) may be employed to delete terminal nucleotides from a DNA fragment to obtain a fragment having a particular desired terminus. Among the commercially available linkers are those that can be ligated to the blunt ends produced by Bal 31 digestion, and which contain restriction endonuclease cleavage site(s). Alternatively, oligonucleotides that reconstruct the N- or C- terminus of a DNA fragment to a desired point may be  
10 synthesized. The oligonucleotide may contain a restriction endonuclease cleavage site upstream of the desired coding sequence and position an initiation codon (ATG) at the N-terminus of the coding sequence.

The gp130 polypeptide is attached to the LIF-R polypeptide through a covalent or non-covalent linkage. Covalent attachment is preferred for certain applications, e.g.  
15 *in vivo* use, in view of the enhanced stability generally conferred by covalent, as opposed to non-covalent, bonds. In constructing the receptor of the present invention, covalent linkage may be accomplished *via* cross-linking reagents, polypeptide linkers, or any other suitable technique.

Numerous reagents useful for cross-linking one protein molecule to another are  
20 known. Heterobifunctional and homobifunctional linkers are available for this purpose from Pierce Chemical Company, Rockford, Illinois, for example. Such linkers contain two functional groups (e.g., esters and/or maleimides) that will react with certain functional groups on amino acid side chains, thus linking one polypeptide to another. The reagent and reaction conditions should be chosen such that the cross-linking does  
25 not interfere with binding of oncostatin M and LIF to the receptor.

One type of polypeptide linker that may be employed in the present invention separates gp130 and LIF-R domains by a distance sufficient to ensure that each domain properly folds into the secondary and tertiary structures necessary for the desired biological activity. The linker also should allow the extracellular domains of gp130 and  
30 LIF-R to assume the proper spatial orientation to form the binding site for oncostatin M and LIF. Suitable polypeptide linkers preferably (1) will adopt a flexible extended conformation, (2) will not exhibit a propensity for developing an ordered secondary structure which could interact with the functional gp130 and LIF-R domains, and (3) will have minimal hydrophobic or charged character which could promote interaction  
35 with the functional protein domains. Typical surface amino acids in flexible protein regions include Gly, Asn and Ser. Virtually any permutation of amino acid sequences containing Gly, Asn and Ser would be expected to satisfy the above criteria for a

peptide linker sequence. Other near neutral amino acids, such as Thr and Ala, may also be used in the linker sequence. Examples of such polypeptide linkers are presented below.

Another type of polypeptide linker that may be employed comprises the Fc region of an antibody. An Fc polypeptide is attached to the C-terminus of LIF-R or of the LIF-R fragment. A separate Fc polypeptide is attached to the C-terminus of gp130 or of the gp130 fragment. The two resulting polypeptide chains may be combined in a buffered solution, whereupon disulfide bonds form between the two Fc polypeptides (e.g., in the so-called hinge region, where interchain disulfide bonds are normally present in antibody molecules). Preferably, a host cell is transformed with DNA encoding both polypeptides such that the two polypeptides are co-expressed and interchain disulfide bonds form in the host cell. LIF-R is thus covalently linked to gp130 via the disulfide bonds in the linker portion of the receptor. Procedures for isolating the Fc region of an antibody are well-known and include proteolytic digestion with papain. Alternatively, an Fc polypeptide may be produced by recombinant cells or chemically synthesized. Also useful are N-terminal fragments of an antibody Fc region that contain the cysteine residues involved in disulfide bond formation at the hinge region. One example of a receptor containing an Fc polypeptide linker is illustrated in example 5 below. The receptor is depicted in Figure 4. The number and position of disulfide bonds may vary from those shown in Figure 4.

Additional examples of LIF-R/Fc and gp130/Fc fusion proteins useful in preparing receptors of the present invention are described in examples 7 and 8. Advantageously, host cells are co-transfected with two different expression vectors, one encoding soluble LIF-R/Fc and the other encoding soluble gp130/Fc. The heterodimer is believed to form intracellularly or during secretion.

Homodimers comprising two LIF-R/Fc polypeptides or two gp130/Fc polypeptides linked via disulfide bonds are also produced by certain of the transfected host cells disclosed herein. The LIF-R/Fc homodimers are useful for binding LIF and the gp130/Fc homodimers find use in binding oncostatin M. The homodimers may be separated from each other and from the heterodimer by virtue of differences in size (e.g., by gel electrophoresis). The heterodimer also may be purified by sequential immunoaffinity chromatography (described below).

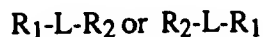
In an alternative embodiment, a first fusion polypeptide comprising gp130 (or fragment thereof) upstream of an antibody light chain (or a fragment thereof) is prepared. A second fusion polypeptide comprises LIF-R upstream of an antibody heavy chain (or a heavy chain fragment, the N-terminus of which extends at least through the C<sub>H</sub>1 region. Disulfide bond(s) form between the gp130-light chain

fusion polypeptide and the LIF-R-heavy chain fusion polypeptide, thus producing a receptor of the present invention comprising a polypeptide linker. If desired, a third fusion (an LIF-R-antibody light chain fusion polypeptide) is prepared and combined with (disulfide bonded to) a fourth fusion comprising gp130 fused to an antibody heavy chain. When the two disulfide bonded molecules are combined, additional disulfide bonds form between the two Fc regions. The resulting receptor of the present invention comprising the four fusion polypeptides resembles an antibody in structure and displays the oncostatin M/LIF binding site bivalently.

A polypeptide linker may be attached to gp130 and to LIF-R by any of the conventional procedures used to attach one polypeptide to another. The cross-linking reagents available from Pierce Chemical Company as described above are among those that may be employed. Amino acids having side chains reactive with such reagents may be included in the polypeptide linker, e.g., at the termini thereof.

The gp130 and LIF-R polypeptides may be separately purified from cellular sources, and then linked together. Alternatively, the receptor of the present invention may be produced using recombinant DNA technology. The gp130 and LIF-R polypeptides may be produced separately and purified from transformed host cells for subsequent covalent linkage. In one embodiment of the present invention, a host cell is transformed/transfected with foreign DNA that encodes gp130 and LIF-R as separate polypeptides. The two polypeptides may be encoded by the same expression vector with start and stop codons for each of the two genes, or the recombinant cells may be co-transfected with two separate expression vectors. In another embodiment, the receptor is produced as a fusion protein in recombinant cells.

In one embodiment of the present invention, the receptor protein is a recombinant fusion protein of the formula:



wherein  $R_1$  represents gp130 or a gp130 fragment;  $R_2$  represents LIF-R or an LIF-R fragment; and L represents a polypeptide linker.

The fusion proteins of the present invention include constructs in which the C-terminal portion of gp130 is fused to the linker which is fused to the N-terminal portion of LIF-R, and also constructs in which the C-terminal portion of LIF-R is fused to the linker which is fused to the N-terminal portion of gp130. gp130 is covalently linked to LIF-R in such a manner as to produce a single protein which retains the desired biological activities of gp130 and LIF-R. The components of the fusion protein are listed in their order of occurrence (i.e., the N-terminal polypeptide is listed first, followed by the linker and then the C-terminal polypeptide).

A DNA sequence encoding a fusion protein is constructed using recombinant DNA techniques to insert separate DNA fragments encoding gp130 and LIF-R into an appropriate expression vector. The 3' end of a DNA fragment encoding gp130 is ligated (*via* the linker) to the 5' end of the DNA fragment encoding LIF-R with the  
5 reading frames of the sequences in phase to permit translation of the mRNA into a single biologically active fusion protein. Alternatively, the 3' end of a DNA fragment encoding LIF-R may be ligated (*via* the linker) to the 5' end of the DNA fragment encoding gp130, with the reading frames of the sequences in phase to permit translation of the mRNA into a single biologically active fusion protein. A DNA sequence  
10 encoding an N-terminal signal sequence may be retained on the DNA sequence encoding the N-terminal polypeptide, while stop codons, which would prevent read-through to the second (C-terminal) DNA sequence, are eliminated. Conversely, a stop codon required to end translation is retained on the second DNA sequence. DNA encoding a signal sequence is preferably removed from the DNA sequence encoding the  
15 C-terminal polypeptide.

Suitable polypeptide linkers comprise a chain of amino acids, preferably from 20 to 100 amino acids in length and most preferably from 30 to 60 amino acids in length. As discussed above, the linker advantageously comprises amino acids selected from the group consisting of glycine, asparagine, serine, threonine, and alanine.  
20 Examples of suitable polypeptide linkers include, but are not limited to,  $(\text{Gly}_4\text{Ser})_n$ , wherein  $n$  is 4-12, preferably 8, and  $(\text{Gly}_4\text{SerGly}_5\text{Ser})_2$ .

A DNA sequence encoding a desired polypeptide linker may be inserted between, and in the same reading frame as, the DNA sequences encoding gp130 and LIF-R using any suitable conventional technique. For example, a chemically  
25 synthesized oligonucleotide encoding the linker and containing appropriate restriction endonuclease cleavage sites may be ligated between the sequences encoding gp130 and LIF-R.

Alternatively, a chemically synthesized DNA sequence may contain a sequence complementary to the 3' terminus (without the stop codon) of either gp130 or LIF-R,  
30 followed by a linker-encoding sequence which is followed by a sequence complementary to the 5' terminus of the other of gp130 and LIF-R. Oligonucleotide directed mutagenesis is then employed to insert the linker-encoding sequence into a vector containing a direct fusion of gp130 and LIF-R.

The present invention provides an isolated DNA sequence encoding the above-described fusion protein comprising gp130, LIF-R, and a polypeptide linker, and also  
35 provides recombinant expression vectors containing the isolated DNA sequence. "Expression vector" refers to a replicable DNA construct used to express DNA which

encodes the desired protein and which includes a transcriptional unit comprising an assembly of (1) genetic element(s) having a regulatory role in gene expression, for example, promoters, operators, or enhancers, operatively linked to (2) a DNA sequence encoding a desired protein (in this case, a receptor of the present invention) which is transcribed into mRNA and translated into protein, and (3) appropriate transcription and translation initiation and termination sequences. The choice of promoter and other regulatory elements generally varies according to the intended host cell.

Proteins to be produced in yeast expression systems preferably include a leader sequence enabling extracellular secretion of translated protein by the yeast host cell. Alternatively, where recombinant protein is expressed without a leader or transport sequence, it may include an N-terminal methionine residue. This residue optionally may be subsequently cleaved from the expressed recombinant protein to provide a final product.

In the expression vectors, regulatory elements controlling transcription or translation are generally derived from mammalian, microbial, viral or insect genes. The ability to replicate in a host, usually conferred by an origin of replication, and a selection gene to facilitate recognition of transformants may additionally be incorporated. Vectors derived from retroviruses also may be employed.

DNA regions are operably linked when they are functionally related to each other. For example, DNA encoding a signal peptide (secretory leader) is operably linked to DNA for a polypeptide if the polypeptide is expressed as a precursor that is secreted through the host cell membrane; a promoter is operably linked to a coding sequence if it controls the transcription of the sequence; or a ribosome binding site is operably linked to a coding sequence if it is positioned so as to permit translation. Generally, "operably linked" means contiguous and, in the case of secretory leaders, contiguous and in reading frame.

Transformed host cells are cells which have been transformed or transfected with foreign DNA using recombinant DNA techniques. In the context of the present invention, the foreign DNA includes a sequence encoding the inventive receptor. Host cells may be transformed for purposes of cloning or amplifying the foreign DNA, or may be transformed with an expression vector for production of the receptor protein. Suitable host cells for expression of the receptor include prokaryotes, yeast or higher eukaryotic cells under the control of appropriate promoters. Prokaryotes include gram negative or gram positive organisms, for example *E. coli* or bacilli. Prokaryotic expression vectors generally comprise one or more phenotypic selectable markers, for example a gene encoding proteins conferring antibiotic resistance or supplying an autotrophic requirement, and an origin of replication recognized by the host to ensure

amplification within the host. Examples of suitable prokaryotic hosts for transformation include *E. coli*, *Bacillus subtilis*, *Salmonella typhimurium*, and various species within the genera *Pseudomonas*, *Streptomyces*, and *Staphylococcus*, although others may also be employed as a matter of choice. Higher eukaryotic cells include  
5 established cell lines of mammalian origin. Cell-free translation systems could also be employed to produce fusion protein using RNAs derived from the DNA constructs of the present invention. Appropriate cloning and expression vectors for use with bacterial, fungal, yeast, and mammalian cellular hosts are described by Pouwels et al. (*Cloning Vectors: A Laboratory Manual*, Elsevier, New York, 1985), the relevant  
10 disclosure of which is hereby incorporated by reference.

Useful expression vectors for bacterial use can comprise a selectable marker and bacterial origin of replication derived from commercially available plasmids comprising genetic elements of the well-known cloning vector pBR322 (ATCC 37017). Such commercial vectors include, for example, pKK223-3 (Pharmacia Fine Chemicals,  
15 Uppsala, Sweden) and pGEM1 (Promega Biotec, Madison, WI, USA). These pBR322 "backbone" sections are combined with an appropriate promoter and the structural sequence to be expressed. *E. coli* is typically transformed using derivatives of pBR322, a plasmid derived from an *E. coli* species (Bolivar et al., *Gene* 2:95, 1977). pBR322 contains genes for ampicillin and tetracycline resistance and this  
20 provides simple means for identifying transformed cells.

Promoters commonly used in recombinant microbial expression vectors include the b-lactamase (penicillinase) and lactose promoter system (Chang et al., *Nature* 275:615, 1978; and Goeddel et al., *Nature* 281:544, 1979), the tryptophan (trp) promoter system (Goeddel et al., *Nucl. Acids Res.* 8:4057, 1980; and EPA 36,776)  
25 and tac promoter (Maniatis, *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory, p. 412, 1982). A particularly useful bacterial expression system employs the phage  $\lambda$  P<sub>L</sub> promoter and cI857ts thermoinducible repressor. Plasmid vectors available from the American Type Culture Collection which incorporate derivatives of the  $\lambda$  P<sub>L</sub> promoter include plasmid pHUB2, resident in *E. coli* strain  
30 JMB9 (ATCC 37092) and pPLc28, resident in *E. coli* RR1 (ATCC 53082).

The recombinant receptor protein may also be expressed in yeast hosts, preferably from *Saccharomyces* species, such as *S. cerevisiae*. Yeast of other genera such as *Pichia* or *Kluyveromyces* may also be employed. Yeast vectors will generally contain an origin of replication from the 2 $\mu$ m yeast plasmid or an autonomously  
35 replicating sequence (ARS), a promoter, DNA encoding the receptor fusion protein, sequences for polyadenylation and transcription termination and a selection gene. Preferably, yeast vectors will include an origin of replication and selectable markers

permitting transformation of both yeast and *E. coli*, e.g., the ampicillin resistance gene of *E. coli* and the *S. cerevisiae* *trp1* gene, which provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan, and a promoter derived from a highly expressed yeast gene to induce transcription of a structural sequence downstream. The presence of the *trp1* lesion in the yeast host cell genome then provides an effective environment for detecting transformation by growth in the absence of tryptophan.

Suitable promoter sequences in yeast vectors include the promoters for metallothionein, 3-phosphoglycerate kinase (Hitzeman et al., *J. Biol. Chem.* 255:2073, 1980) or other glycolytic enzymes (Hess et al., *J. Adv. Enzyme Reg.* 7:149, 1968; and Holland et al., *Biochem.* 17:4900, 1978), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase and glucokinase. Suitable vectors and promoters for use in yeast expression are further described in R. Hitzeman et al., EPA 73,657.

Preferred yeast vectors can be assembled using DNA sequences from pBR322 for selection and replication in *E. coli* (*Amp<sup>r</sup>* gene and origin of replication) and yeast DNA sequences including a glucose-repressible ADH2 promoter and  $\alpha$ -factor secretion leader. The ADH2 promoter has been described by Russell et al. (*J. Biol. Chem.* 258:2674, 1982) and Beier et al., (*Nature* 300:724, 1982). The yeast  $\alpha$ -factor leader, which directs secretion of heterologous proteins, can be inserted between the promoter and the structural gene to be expressed. See, e.g., Kurjan et al., *Cell* 30:922, 1982; and Bitter et al., *Proc. Natl. Acad. Sci. USA* 81:5330, 1984. The leader sequence may be modified to contain, near its 3' end, one or more useful restriction sites to facilitate fusion of the leader sequence to foreign genes.

Suitable yeast transformation protocols are known to those of skill in the art. An exemplary technique is described by Hinnen et al., *Proc. Natl. Acad. Sci. USA* 75:1929, (1978), selecting for *Trp<sup>+</sup>* transformants in a selective medium consisting of 0.67% yeast nitrogen base, 0.5% casamino acids, 2% glucose, 10  $\mu$ g/ml adenine and 20  $\mu$ g/ml uracil.

Host strains transformed by vectors comprising the ADH2 promoter may be grown for expression in a rich medium consisting of 1% yeast extract, 2% peptone, and 1% glucose supplemented with 80  $\mu$ g/ml adenine and 80  $\mu$ g/ml uracil. Derepression of the ADH2 promoter occurs upon exhaustion of medium glucose. Crude yeast supernatants are harvested by filtration and held at 4°C prior to further purification.

Various mammalian or insect cell culture systems can be employed to express recombinant protein. Baculovirus systems for production of heterologous proteins in insect cells are reviewed by Luckow and Summers, *Bio/Technology* 6:47 (1988). Examples of suitable mammalian host cell lines include the COS-7 lines of monkey  
5 kidney cells, described by Gluzman (*Cell* 23:175, 1981), and other cell lines capable of expressing an appropriate vector including, for examples, L cells, C127, 3T3, Chinese hamster ovary (CHO), HeLa and BHK cell lines. Mammalian expression vectors may comprise non-transcribed elements such as an origin of replication, a suitable promoter and enhancer linked to the gene to be expressed, and other 5' or 3' flanking  
10 nontranscribed sequences, and 5' or 3' nontranslated sequences, such as necessary ribosome binding sites, a poly-adenylation site, splice donor and acceptor sites, and transcriptional termination sequences.

The transcriptional and translational control sequences in expression vectors to be used in transforming vertebrate cells may be provided by viral sources. For  
15 example, commonly used promoters and enhancers are derived from Polyoma, Adenovirus 2, Simian Virus 40 (SV40), and human cytomegalovirus. DNA sequences derived from the SV40 viral genome, for example, SV40 origin, early and late promoter, enhancer, splice, and polyadenylation sites may be used to provide the other genetic elements required for expression of a heterologous DNA sequence. The early  
20 and late promoters are particularly useful because both are obtained easily from the virus as a fragment which also contains the SV40 viral origin or replication (Fiers et al., *Nature* 273:113, 1978). Smaller or larger SV40 fragments may also be used, provided the approximately 250 bp sequence extending from the Hind III site toward the *Bgl*II site located in the viral origin of replication is included. Exemplary vectors can be  
25 constructed as disclosed by Okayama and Berg (*Mol. Cell. Biol.* 3:280, 1983).

A useful system for stable high level expression of mammalian receptor cDNAs in C127 murine mammary epithelial cells can be constructed substantially as described by Cosman et al. (*Mol. Immunol.* 23:935, 1986).

Particularly preferred vectors for expression of the inventive receptor as a  
30 fusion protein are described in the examples below. The foregoing discussion is, of course, applicable to the production of recombinant fusion proteins comprising a fragment of gp130 and/or a fragment of LIF-R. Suitable fragments are discussed above, and DNA sequences encoding such fragments may be inserted into the above-described expression vectors.

35 The present invention provides a process for preparing the recombinant receptor of the present invention, comprising culturing a host cell transformed with an expression vector comprising a DNA sequence that encodes said receptor under

conditions that promote expression. The receptor is then purified from culture media or cell extracts.

For example, supernatants from systems which secrete recombinant protein into the culture medium can be first concentrated using a commercially available protein concentration filter, for example, an Amicon or Millipore Pellicon ultrafiltration unit. Following the concentration step, the concentrate can be applied to a suitable purification matrix. For example, a suitable affinity matrix can comprise LIF or OSM. An LIF affinity matrix may be prepared by coupling recombinant human LIF to cyanogen bromide-activated Sepharose (Pharmacia) or Hydrazide Affigel (Biorad), according to manufacturer's recommendations. Sequential immunopurification using antibodies bound to a suitable support is preferred. Proteins binding to an antibody specific for LIF-R are recovered and contacted with antibody specific for gp130 on an insoluble support. Proteins immunoreactive with both antibodies may thus be identified and isolated. Alternatively, an anion exchange resin can be employed, for example, a matrix or substrate having pendant diethylaminoethyl (DEAE) groups. The matrices can be acrylamide, agarose, dextran, cellulose or other types commonly employed in protein purification. Alternatively, a cation exchange step can be employed. Suitable cation exchangers include various insoluble matrices comprising sulfopropyl or carboxymethyl groups. Sulfopropyl groups are preferred. One or more reversed-phase high performance liquid chromatography (RP-HPLC) steps employing hydrophobic RP-HPLC media, e.g., silica gel having pendant methyl or other aliphatic groups, can be employed to further purify a fusion protein composition.

Some or all of the foregoing purification steps, in various combinations, can be employed to provide an essentially homogeneous recombinant protein. Recombinant cell culture enables the production of the fusion protein free of those contaminating proteins which may be normally associated with gp130 or LIF-R as they are found in nature in their respective species of origin, e.g., in cells, cell exudates or body fluids.

The foregoing purification procedures are among those that may be employed to purify non-recombinant receptors of the present invention as well. When linking procedures that may produce homodimers (gp130-linker-gp130 and LIF-R-linker-LIF-R) are employed, purification procedures that separate the desired heterodimer from such homodimers are employed. An example of such a procedure is sequential immunopurification as discussed above.

Recombinant protein produced in bacterial culture is usually isolated by initial extraction from cell pellets, followed by one or more concentration, salting-out, aqueous ion exchange or size exclusion chromatography steps. Finally, high performance liquid chromatography (HPLC) can be employed for final purification

steps. Microbial cells employed in expression of recombinant fusion proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents.

5 Fermentation of yeast which express fusion proteins as a secreted protein greatly simplifies purification. Secreted recombinant protein resulting from a large-scale fermentation can be purified by methods analogous to those disclosed by Urdal et al. (*J. Chromatog.* 296:171, 1984), involving two sequential, reversed-phase HPLC steps for purification of a recombinant protein on a preparative HPLC column.

10 The present invention also provides a pharmaceutical composition comprising a receptor protein of the present invention with a physiologically acceptable carrier or diluent. Such carriers and diluents will be nontoxic to recipients at the dosages and concentrations employed. Such compositions may, for example, comprise the receptor protein in a buffered solution, to which may be added antioxidants such as ascorbic acid, low molecular weight (less than about ten residues) polypeptides, proteins, amino acids, carbohydrates including glucose, sucrose or dextrans, chelating agents such as EDTA, glutathione and other stabilizers and excipients. The receptor of the present invention may be administered by any suitable method in a manner appropriate to the indication, such as intravenous injection, continuous infusion, sustained release from implants, etc.

20 The DNA and/or amino acid sequences of gp130 and LIF-R may vary from those presented in SEQ ID NO: 1 and SEQ ID NO: 5. Due to the known degeneracy of the genetic code, there can be considerable variation in nucleotide sequences encoding the same amino acid sequence. The DNA sequences capable of hybridizing to the native DNA sequence of SEQ ID NO: 1 or SEQ ID NO: 5 under moderately stringent conditions (50°C, 2 X SSC), and which encode a biologically active gp130 or LIF-R polypeptide, are also considered to be gp130-encoding or LIF-R-encoding DNA sequences, respectively, in the context of the present invention. Further, certain mutations in a nucleotide sequence which encodes LIF-R or gp130 will not be expressed in the final protein product. For example, nucleotide substitutions may be made to enhance expression, primarily to avoid secondary structure loops in the transcribed mRNA (see EPA 75,444A, incorporated herein by reference). Other alterations of the nucleotide sequence may be made to provide codons that are more readily translated by the selected host, e.g., the well-known *E. coli* preference codons for *E. coli* expression.

35 The amino acid sequence of native gp130 or LIF-R may be varied by substituting, deleting, adding, or inserting one or more amino acids to produce a gp130 or LIF-R variant. Variants that possess the desired biological activity of the native

gp130 and LIF-R proteins may be employed in the receptor of the present invention. Assays by which the biological activity of variant proteins may be analyzed are described in the examples below.

Alterations to the native amino acid sequence may be accomplished by any of a number of known techniques. For example, mutations can be introduced at particular loci by synthesizing oligonucleotides containing a mutant sequence, flanked by restriction sites enabling ligation to fragments of the native sequence. Following ligation, the resulting reconstructed sequence encodes an analog having the desired amino acid insertion, substitution, or deletion.

Alternatively, oligonucleotide-directed site-specific mutagenesis procedures can be employed to provide an altered gene having particular codons altered according to the substitution, deletion, or insertion required. Exemplary methods of making the alterations set forth above are disclosed by Walder et al. (*Gene* 42:133, 1986); Bauer et al. (*Gene* 37:73, 1985); Craig (*BioTechniques*, January 1985, 12-19); Smith et al. (*Genetic Engineering: Principles and Methods*, Plenum Press, 1981); U.S. Patent No. 4,518,584, and U.S. Patent No. 4,737,462, which are incorporated by reference herein.

Bioequivalent variants of LIF-R and gp130 may be constructed by, for example, making various substitutions of amino acid residues or deleting terminal or internal amino acids not needed for biological activity. In one embodiment of the invention, the variant amino acid sequence is at least 80% identical, preferably at least 90% identical, to the native sequence. Percent similarity may be determined, for example, by comparing sequence information using the GAP computer program, version 6.0, available from the University of Wisconsin Genetics Computer Group (UWGCG). The GAP program utilizes the alignment method of Needleman and Wunsch (*J. Mol. Biol.* 48:443, 1970), as revised by Smith and Waterman (*Adv. Appl. Math.* 2:482, 1981). Briefly, the GAP program defines similarity as the number of aligned symbols (i.e., nucleotides or amino acids) which are similar, divided by the total number of symbols in the shorter of the two sequences. The preferred default parameters for the GAP program include: (1) a unary comparison matrix (containing a value of 1 for identities and 0 for non-identities) for nucleotides, and the weighted comparison matrix of Gribskov and Burgess, *Nucl. Acids Res.* 14:6745, 1986, as described by Schwartz and Dayhoff, eds., *Atlas of Protein Sequence and Structure*, National Biomedical Research Foundation, pp. 353-358, 1979; (2) a penalty of 3.0 for each gap and an additional 0.10 penalty for each symbol in each gap; and (3) no penalty for end gaps.

Generally, substitutions should be made conservatively; i.e., the most preferred substitute amino acids are those having physiochemical characteristics resembling those of the residue to be replaced. Examples of conservative substitutions include substitution of one aliphatic residue for another, such as Ile, Val, Leu, or Ala for one  
5 another, or substitutions of one polar residue for another, such as between Lys and Arg; Glu and Asp; or Gln and Asn. Other such conservative substitutions, for example, substitutions of entire regions having similar hydrophobicity characteristics, are well known. Moreover, particular amino acid differences between human, murine and other mammalian LIF-Rs is suggestive of additional conservative substitutions that  
10 may be made without altering the essential biological characteristics of LIF-R.

Cysteine residues can be deleted or replaced with other amino acids to prevent formation of unnecessary or incorrect intramolecular disulfide bridges upon renaturation. Adjacent dibasic amino acid residues may be modified to enhance expression in yeast systems in which KEX2 protease activity is present.

15 EP212,914 discloses the use of site-specific mutagenesis to inactivate KEX2 protease processing sites in a protein. KEX2 protease processing sites are inactivated by deleting, adding or substituting residues to alter Arg-Arg, Arg-Lys, and Lys-Arg pairs to eliminate the occurrence of these adjacent basic residues. Lys-Lys pairings are considerably less susceptible to KEX2 cleavage, and conversion of Arg-Lys or Lys-  
20 Arg to Lys-Lys represents a conservative and preferred approach to inactivating KEX2 sites.

Hydrophilic amino acids may be substituted for hydrophobic amino acids in the transmembrane region and/or intracellular domain of gp130 and LIF-R to enhance water solubility of the proteins. Addition of amino acids to the native sequence may  
25 result from translation of in-frame codons present in linkers used in constructing cloning or expression vectors. The LIF-R encoded by clone pHLIF-R-65 contains such linker-encoded amino acids at the C-terminus, as described by Gearing et al., *supra*.

The present invention also includes proteins with or without associated native-  
30 pattern glycosylation. Expression of DNAs encoding the fusion proteins in bacteria such as *E. coli* provides non-glycosylated molecules. Functional mutant analogs having inactivated N-glycosylation sites can be produced by oligonucleotide synthesis and ligation or by site-specific mutagenesis techniques. These analog proteins can be produced in a homogeneous, reduced-carbohydrate form in good yield using yeast  
35 expression systems. N-glycosylation sites in eukaryotic proteins are characterized by the amino acid triplet Asn-A<sub>1</sub>-Z, where A<sub>1</sub> is any amino acid except Pro, and Z is Ser or Thr. In this sequence, asparagine provides a side chain amino group for covalent

attachment of carbohydrate. Such a site can be eliminated by substituting another amino acid for Asn or for residue Z, deleting Asn or Z, or inserting a non-Z amino acid between A<sub>1</sub> and Z, or an amino acid other than Asn between Asn and A<sub>1</sub>. Known procedures for inactivating N-glycosylation sites in proteins include those described in U.S. Patent 5,071,972 and EP 276,846.

5 Variants of the receptor proteins of the present invention also include various structural forms of the primary protein which retain biological activity. Due to the presence of ionizable amino and carboxyl groups, for example, a receptor protein may be in the form of acidic or basic salts, or may be in neutral form. Individual amino acid residues may also be modified by oxidation or reduction.

10 The primary amino acid structure also may be modified by forming covalent or aggregative conjugates with other chemical moieties, such as glycosyl groups, lipids, phosphate, acetyl groups and the like. Covalent derivatives are prepared by linking particular functional groups to amino acid side chains or at the N- or C- termini. Other derivatives of the receptor protein within the scope of this invention include covalent or aggregative conjugates of the receptor protein with other proteins or polypeptides, such as by synthesis in recombinant culture as N- or C- terminal fusions. For example, the conjugated polypeptide may be a signal (or leader) polypeptide sequence at the N-terminal region of the protein which co-translationally or post-translationally directs transfer of the protein from its site of synthesis to its site of function inside or outside of the cell membrane or wall (e.g., the yeast *a-factor* leader). Peptides may also be added to facilitate purification or identification of the fusion protein (e.g., poly-His). The amino acid sequence of the fusion protein can also be linked to the peptide Asp-Tyr-Lys-Asp-Asp-Asp-Asp-Lys (DYKDDDDK) (Hopp et al., *Bio/Technology* 6:1204, 1988). The latter sequence is highly antigenic and provides an epitope reversibly bound by a specific monoclonal antibody, enabling rapid assay and facile purification of expressed recombinant protein. This sequence is also specifically cleaved by bovine mucosal enterokinase at the residue immediately following the Asp-Lys pairing. Receptor proteins capped with this peptide may also be resistant to intracellular degradation in *E. coli*.

30 The receptors of the present invention are primarily useful as oncostatin M binding reagents, and may be administered *in vivo* to inhibit a biological activity of oncostatin M (including signal transduction). The inventive receptors also have use as LIF binding reagents.

35 Disorders mediated by either oncostatin M or LIF may be treated by administering a therapeutically effective amount of the receptor of the present invention to a human or mammalian patient afflicted with such a disorder. A disorder is said to

be mediated by oncostatin M or LIF when biologically active oncostatin M or LIF causes (directly or indirectly) or exacerbates the disorder. Soluble receptor proteins can be used to competitively bind to LIF and oncostatin M, thereby inhibiting binding of LIF and oncostatin M to cell surface receptors.

5           As discussed in example 2, gp130 has now been found to bind oncostatin M, although with lower affinity than the inventive receptors comprising both gp130 and LIF-R. gp130 may be administered to treat conditions mediated by oncostatin M, although a gp130/LIF-R receptor of the present invention would be preferred for such a purpose.

10           Oncostatin M has been reported to stimulate hematopoiesis, stimulate epithelial cell proliferation, increase plasmin activity (thereby inducing fibrinolysis), inhibit angiogenesis and suppress expression of major histocompatibility complex antigens on endothelial cells. See PCT application WO 9109057 and European patent application no. 422,186. When these or other biological effects of oncostatin M are undesirable, a  
15           receptor of the present invention may be administered to bind oncostatin M.

          Oncostatin M is believed to stimulate production of the cytokine interleukin-6 (IL-6), as reported by Brown et al., *J. Immunol.* 147:2175 (1991). Oncostatin M therefore indirectly mediates disorders associated with the presence of IL-6. IL-6 has been reported to be involved in the pathogenesis of AIDS-associated Kaposi's sarcoma  
20           (deWit et al., *J. Intern. Med. [England]* 229:539 [1991]). Binding of oncostatin M by a receptor of the present invention thus may be useful in treating Kaposi's sarcoma. Alternatively, but less preferably, gp130 may be administered to treat Kaposi's sarcoma.

          Among the disorders mediated by LIF are lipoprotein metabolism defects such  
25           as atherosclerosis and obesity, as well as disorders of bone and calcium metabolism or disorders associated with LIF overproduction that affect hepatocytes, neurons, or leukocytes. The regulation of embryonic and hematopoietic stem cells by LIF may also be manipulated with the receptor. A soluble form of the receptor may also be used to treat leukemic cells which respond to LIF by proliferating. LIF also may play a role in  
30           inducing cachexia in cancer or AIDS patients. The receptor, or antibodies thereto, may also be useful as a diagnostic reagent to detect diseases characterized by the presence of abnormal LIF-R.

          Oncostatin M and LIF are different proteins, but share certain structural and biological properties. If inhibition of a biological activity shared by oncostatin M and  
35           LIF is desired, the receptor of the present invention offers the benefit of binding both of these proteins exhibiting the particular biological activity. A receptor binding only one

of the proteins would leave the other protein active and continuing to mediate the disorder.

Receptor proteins or derivatives thereof may also be used as reagents in receptor-based immunoassays, reagents in assays for oncostatin M or LIF, or as binding agents for affinity purification of oncostatin M or LIF. The receptor proteins of the present invention may be used as immunogens in conventional procedures for production of polyclonal or monoclonal antibodies. Such antibodies may be employed on immunoaffinity columns for purification of the receptor, or as components of diagnostic or research assays. Derivatives may also be obtained by attacking additional polypeptide(s), e.g., by using a cross-linking agent, such as N-maleimidobenzoyl succinimide ester that reacts with cysteine and lysine residues. Receptor proteins may also be covalently bound through reactive side groups to various insoluble substrates, such as cyanogen bromide-activated, bisoxirane-activated, carbonyldiimidazole-activated or tosyl-activated agarose structures, or by adsorbing to polyolefin surfaces (with or without glutaraldehyde cross-linking).

The following examples are provided to illustrate certain embodiments of the invention, and are not to be construed as limiting the scope of the invention.

## EXAMPLES

20

### Example 1

#### Assay to Detect Binding of LIF

Recombinant human LIF is expressed in yeast and purified to homogeneity essentially as described by Hopp, et al., *Bio/Technology* 6:1204 (1988). The purified protein is radiolabeled using a commercially available enzymobead radioiodination reagent (BioRad). In this procedure 10 µg LIF in 50 µl 0.2 M sodium phosphate, pH 7.2, are combined with 50 µl enzymobead reagent, 2 mCi of sodium iodide in 20 µl of 0.05 M sodium phosphate pH 7.0 and 10 µl of 2.5% β-D-glucose. After 10 minutes at 25°C, sodium azide (20 µl of 50 mM) and sodium metabisulfite (10 µl of 5 mg/ml) are added and incubation is continued for 5 min. at 25°C. The reaction mixture is fractionated by gel filtration on a 2 ml bed volume of Sephadex® G-25 (Sigma) equilibrated in Roswell Park Memorial Institute (RPMI) 1640 medium containing 2.5% (w/v) bovine serum albumin (BSA), 0.2% (w/v) sodium azide and 20 mM Hepes pH 7.4 (binding medium). The final pool of <sup>125</sup>I-LIF is diluted to a working stock solution of 3 x 10<sup>-8</sup> M in binding medium and stored for up to one month at 4°C

without detectable loss of receptor binding activity. The specific activity is routinely in the range of  $6-8 \times 10^{15}$  cpm/mmol LIF.

5 The radiolabeled LIF may be employed in any of a number of conventional assay procedures to determine whether a given protein or cell binds LIF. Examples of such assays are those that detect binding of the radiolabeled LIF to cells expressing an LIF-binding protein on the cell surface. The radiolabeled LIF also may be employed in assays for the presence of LIF-binding proteins in cell culture medium (e.g. LIF-binding proteins secreted by recombinant cells). Proteins in cell extracts (e.g. from recombinant cells) also may be assayed for the ability to bind the radiolabeled LIF.

10 In one assay procedure, cells transformed/transfected with an expression system encoding a protein to be tested for ability to bind LIF are plated at a density of  $2 \times 10^5$  cells/well in either 6 well dishes (Falcon) or single well chambered slides (Lab-Tek). Both dishes and slides are pretreated with 1 ml human fibronectin (10  $\mu$ g/ml in PBS) for 30 minutes followed by 1 wash with PBS. After 48 to 72 hours, cells are  
15 assayed for expression of LIF binding proteins by binding radioiodinated LIF using the following slide autoradiography technique. Transfected cells are washed once with binding medium (RPMI media 1640 containing 25 mg/ml bovine serum albumin (BSA), 2 mg/ml sodium azide, 20 mM HEPES, pH 7.2, and 50 mg/ml nonfat dry milk (NFDM) and incubated for 2 hours at 4°C with 1 ml binding medium + NFDM  
20 containing  $1.25 \times 10^{-9}$  M  $^{125}$ I-LIF. After incubation, cells in the chambered slides are washed three times with binding buffer + NFDM, followed by 2 washes with PBS, pH 7.3, to remove unbound  $^{125}$ I-LIF. The cells are fixed by incubating for 30 minutes at room temperature in 10% glutaraldehyde in PBS, pH 7.3, washed twice in PBS, and air dried. The slides are dipped in Kodak NTB-2 photographic emulsion (5x dilution in water) and exposed in the dark for 12 hours to 7 days at 4°C in a light proof box. The  
25 slides are then developed for approximately 5 minutes in Kodak D19 developer (40 g/500 ml water), rinsed in water and fixed in Agfa G433C fixer. The slides are individually examined with a microscope at 25-40x magnification and positive cells that bind LIF are identified by the presence of autoradiographic silver grains against a light  
30 background.

Cells in the 6 well plates are washed once with binding buffer + NFDM followed by 3 washings with PBS, pH 7.3, to remove unbound  $^{125}$ I-LIF. The cells are then trypsinized to remove them from the plate and bound  $^{125}$ I-LIF is counted on a gamma counter.

35 The cells in transfectant pool(s) testing positive are subdivided into smaller pools and the screening process is repeated (with further subdividing of the pools as necessary) until an individual clone expressing LIF-binding protein is isolated. Non-

specific binding of  $^{125}\text{I}$ -LIF may be measured in the presence of 200-fold or greater excess of unlabeled LIF. As a control, the same host cells transfected with a vector lacking LIF-R-encoding sequences should be assayed to determine whether background endogenous LIF receptors are present on the host cells.

5 In another assay procedure, cells producing a soluble LIF-binding protein that is released from the cells into the culture medium may be identified. Cells are collected by centrifugation from a culture broth. The supernatant (culture medium) is concentrated 10-fold, and 1  $\mu\text{l}$  aliquots are spotted onto nitrocellulose filters and allowed to air dry. Additional binding sites are blocked by overnight incubation at 4°C  
10 in the above-described binding medium containing 3% non-fat dry milk (BMNFDM). Filters are incubated for 2h at 4°C in BMNFDM containing 1 nM  $^{125}\text{I}$ -LIF in the presence or absence of 200 nM unlabeled LIF, then washed (3 x 5 min) in PBS. Filters are exposed to photographic film for 48 hr at room temperature.

The results of one LIF binding assay conducted according to the following  
15 procedure are shown in Figure 1. Host cells transfected with vector(s) encoding LIF-R or gp130 as described below were assayed for the ability to bind LIF. The host cells were the monkey kidney cell line designated COS-7, described by Glutzman, *Cell* 23:175 (1981). In separate transfections, COS-7 cells were transfected with the following combinations of vectors. The different types of transfected cells (and non-  
20 transfected control cells) are designated A-F as shown below, and the curves representing the LIF-binding assay data for each transfected or control cell type are also labeled A-F in Figure 1.

(A) B10G (the gp130 encoding vector described in example 3) and  
25 pHLIFR-65 (the LIF-R encoding vector described in example 3)

(B) pHLIFR-65 and control vector CAV (a control vector that does not  
encode LIF-R or gp130; controls for plasmid dilution so that results can be more  
accurately compared with those of COS-7 cells co-transfected with both a gp130  
30 encoding vector and an LIF-R encoding vector)

(C) B10G and pHLIFR-65; transfected cells were preincubated with  
non-radiolabeled oncostatin M before incubation with  $^{125}\text{I}$ -LIF

35 (D) pHLIFR-65 and CAV; transfected cells were preincubated with non-  
radiolabeled oncostatin M before incubation with  $^{125}\text{I}$ -LIF

(E) non-transfected COS-7 cells (control)

(F) B10G and CAV

5       The assay was performed by a phthalate oil separation method essentially as described by Dower et al., *J. Immunol.* 132:751 (1984) and Park et al., *J. Biol. Chem.* 261:4177 (1986). Briefly, the COS-7 host cells were released from 10 cm tissue culture plates two days after transfection by incubation in non-enzymatic cell dissociation buffer (Sigma) at 37°C for 30-60 minutes. Cells were then washed with  
10   the above-described binding medium and resuspended in binding medium at  $5 \times 10^6$  cells/ml. 50 $\mu$ l aliquots of the cells were incubated with serial dilutions of  $^{125}$ I-LIF at room temperature for one hour with agitation (in the presence or absence of a 200-fold excess of unlabeled LIF) in a total volume of 150 $\mu$ l. The unlabeled LIF allowed for calculation of the non-specific background binding of LIF. Duplicate aliquots (60 $\mu$ l) of  
15   each incubation mixture were then transferred to a polyethylene centrifuge tube containing a phthalate oil mixture comprising 1.5 parts dibutylphthalate to 1 part bis(s-ethylhexyl)phthalate.

      The cells were separated from unbound  $^{125}$ I-LIF by centrifugation for five minutes at 15,000 x g in an Eppendorf microfuge. The centrifuge tubes were cut to  
20   separate the pellet of cells (containing bound  $^{125}$ I-LIF) from the supernatant containing unbound  $^{125}$ I-LIF. The radioactivity in both parts was then determined on a gamma counter. The determinations of both cell-bound and unbound radioactivity from the two 60 $\mu$ l aliquots were averaged for subsequent calculations.

      The results are presented in Figure 1 as standard Scatchard transformations of  
25   the biological data. The data are reported as the ratio of molecules of  $^{125}$ I-LIF bound per cell, to free  $^{125}$ I-LIF molecules (y-axis) versus molecules of  $^{125}$ I-LIF bound per cell (x-axis). The dissociation constants ( $K_D$ ) are shown in Figure 1, along with the number of LIF-binding sites per cell. Since a saturating amount of radiolabeled LIF was offered, the number of molecules of radiolabeled LIF bound per cell is considered  
30   equivalent to the number of LIF binding sites per cell.

      As shown by curve A of Figure 1, COS-7 cells co-transfected with a gp130 encoding vector (B10G) and an LIF-R encoding vector (pHLIFR-65) demonstrated high affinity LIF binding ( $K_D=9 \times 10^{-10}$ M). When these same co-transfected COS-7 cells were preincubated with non-radiolabeled oncostatin M before incubation with  
35    $^{125}$ I-LIF (curve C), binding of LIF was greatly reduced ( $K_D=4.2 \times 10^{-9}$ M). Oncostatin M thus competes with LIF for binding sites on these transfected cells.

COS-7 cells transfected with a vector encoding the single-polypeptide chain LIF-R (pHLIFR-65) and with the control vector CAV bound LIF (curve B;  $K_D=2.4 \times 10^{-9}M$ ), but with lower affinity than the cells producing both gp130 and LIF-R. The COS-7 cells display endogenous high affinity simian LIF receptors (curve E;  $K_D$  about  $3 \times 10^{-11}M$ ). Transfection with pHLIFR-65 (encoding the single polypeptide LIF-R) results in display of additional low affinity LIF receptors ( $K_D=2.4 \times 10^{-9}M$ ; curve B, site 2) as well as the simian LIF receptors  $K_D=3.3 \times 10^{-11}M$ ; curve B, site 1).

When the COS-7 cells transfected with pHLIFR-65 and CAV were preincubated with non-radiolabeled oncostatin M before incubation with  $^{125}I$ -LIF (CURVE D), binding of LIF to the LIF-R expressed by pHLIFR-65 was essentially unchanged compared to the same transfected cells not preincubated with oncostatin M. Oncostatin M thus does not compete with LIF for binding to the single polypeptide chain LIF-R. However, the binding of LIF to the endogenous simian high affinity LIF-R on the COS-7 cells was competed.

The COS-7 cells co-transfected with the gp130 encoding vector and the CAV control vector (curve F) did not bind LIF in any measurable amount above the amount of binding to the non-transfected COS-7 cells (curve E).

## Example 2

### Assay to Detect Binding of Oncostatin M

Oncostatin M may be purified from cells in which the protein is naturally found, or from cells transformed with an expression vector encoding oncostatin M. One source of oncostatin M is phorbol ester-treated U937 cells, as described by Zarling et al., *PNAS U.S.A.* 83:9739 (1986). Purification of recombinant oncostatin M is described by Linsley et al., *J. Biol. Chem.* 264:4282-4289 (1989), which is hereby incorporated by reference in its entirety.

Preferably, oncostatin M is produced in yeast cells transformed with a suitable expression vector. A DNA sequence encoding a signal sequence (e.g., a yeast alpha-factor leader sequence) may be fused to the N-terminus of the oncostatin M encoding DNA sequence to promote secretion of the protein from the host cells. The protein when initially produced may also comprise an N-terminal identification leader (e.g., a "flag" sequence such as Asp-Tyr-Lys-Asp<sub>4</sub>-Lys) as described by Hopp et al., *BioTechnology* 6:1204 (1988). The flag sequence is highly antigenic and provides an epitope reversibly bound by a specific monoclonal antibody, enabling facile purification of the expressed recombinant protein. This sequence is also specifically cleaved by bovine mucosal enterokinase at the residue immediately following the Asp-Lys pairing.

Neither the signal sequence nor the flag sequence is found on the processed final oncostatin M product.

Oncostatin M may be radiolabeled using any suitable conventional procedure, such as the radioiodination procedure employed to radiolabel LIF in Example 1. The  
5 radio-iodination of oncostatin M has also been described by Linsley et al., *supra*.

The resulting radiolabeled oncostatin M may be substituted for radiolabeled LIF (using the same concentrations and other reaction parameters) in the assay procedures described in Example 1 in order to detect proteins and cells that bind oncostatin M. An assay for binding of  $^{125}\text{I}$ -oncostatin M to cells is also described in Linsley et al.,  
10 *supra*.

The results of one oncostatin M binding assay are shown in Figure 2. COS-7 cells transfected with vector(s) encoding gp130 or LIF-R were assayed for the ability to bind oncostatin M. In separate transfections, COS-7 cells were transfected with the following combinations of vectors. The different types of transfected cells (and non-  
15 transfected control cells) are designated A-E as shown below, and the corresponding curves representing the oncostatin M binding assay data for each cell type are also labeled A-E in Figure 2.

(A) B10G (the gp130 encoding vector described in example 3) and  
20 pHLIFR-65 (the LIF-R encoding vector described in example 3)

(B) B10G and pHLIFR-65; transfected cells were preincubated with non-radiolabeled LIF before incubation with  $^{125}\text{I}$ -oncostatin M

25 (C) pHLIFR-65 and CAV (a control vector that does not encode LIF-R or gp130; controls for plasmid dilution so that results can be more accurately compared with those of COS-7 cells co-transfected with both a gp130 encoding vector and an LIF-R encoding vector)

30 (D) non-transfected COS-7 cells (control)

(E) B10G and CAV

The assay was performed by the phthalate oil separation method described in  
35 example 1 (but substituting oncostatin M for LIF). The results are presented in Figure 2 as standard Scatchard transformations of the biological data. The data are reported as the ratio of molecules of  $^{125}\text{I}$ -oncostatin M bound per cell, to free  $^{125}\text{I}$ -oncostatin M

molecules (y-axis) versus molecules of  $^{125}\text{I}$ -oncostatin M bound per cell (x-axis). The dissociation constants ( $K_D$ ) are shown in Figure 2, along with the number of oncostatin M-binding sites per cell. Since a saturating amount of radiolabeled oncostatin M was offered, the number of molecules of radiolabeled oncostatin M bound per cell is  
5 considered equivalent to the number of oncostatin M binding sites per cell.

As shown by curve A in Figure 2, COS-7 cells co-transfected with a gp130 encoding vector (B10G) and an LIF-R encoding vector (pHLIFR-65) demonstrated the ability to bind oncostatin M with high affinity ( $K_D=2.4 \times 10^{-10}\text{M}$ ).

COS-7 cells co-transfected with a vector encoding the single-polypeptide chain  
10 LIF-R (pHLIFR-65) and with the control vector CAV (curve C) did not bind oncostatin M in any significant amount above that bound by the non-transfected COS-7 cells (curve D).

COS-7 cells co-transfected with pHLIFR-65 and B10G and preincubated with non-radiolabeled LIF before incubation with  $^{125}\text{I}$ -oncostatin M (curve B) did not bind  
15 oncostatin M in any measurable amount above that bound by the non-transfected COS-7 cells. LIF thus competes with oncostatin M for binding sites on the recombinant cells.

The experimental conditions of this assay (the results of which are shown in Figure 2) were not appropriate for accurate detection of low affinity oncostatin M  
20 receptors. Thus, a separate experiment (phthalate oil separation method) was conducted to compare oncostatin M binding by COS-7 cells transfected with B10G alone (no CAV control vector) with oncostatin M binding to non-transfected COS-7 cells. Non-transfected COS-7 cells assayed as a control demonstrated a small number of high affinity oncostatin M receptors ( $K_D=3.6 \times 10^{-10}\text{M}$ ). The cells transfected with  
25 B10G demonstrated additional low affinity binding of oncostatin M ( $K_D=7.7 \times 10^{-9}\text{M}$ ). The results of this oncostatin M binding assay are shown in Figure 3 as Scatchard transformations of the biological data. The data are reported as the ratio of molecules of  $^{125}\text{I}$ -oncostatin M bound per cell, to free  $^{125}\text{I}$ -oncostatin M molecules (y-axis) versus molecules of  $^{125}\text{I}$ -oncostatin M bound per cell (x-axis). The scale in Figure 3 differs  
30 from that of Figures 1 and 2 so that the difference in oncostatin M binding by the gp130-producing cells compared to the control cells can be more readily visualized.

Disorders mediated by oncostatin M thus may be treated by administering gp130 or a fragment thereof. Receptors comprising both gp130 and LIF-R are preferred for use in treating such conditions, however, in view of the higher affinity of  
35 such receptors for oncostatin M compared to the affinity of gp130 alone for oncostatin M. gp130 also may be employed as an oncostatin M-binding reagent in diagnostic and research assays.

**Example 3****Preparation of a Recombinant Fusion Protein Designated LIF-R-Linker-gp130**

5 A recombinant receptor protein of the present invention is prepared by the following procedure. The receptor comprises an LIF-R fragment at the N-terminus attached to a gp130 fragment through a polypeptide linker. The polypeptide linker is of the formula (Gly<sub>4</sub>Ser)<sub>8</sub>. An oligonucleotide encoding a portion of the linker sequence, i.e., the sequence Ser(Gly<sub>4</sub>Ser)<sub>6</sub>Gly is synthesized by any of the conventional known  
 10 procedures for oligonucleotide synthesis. The DNA and encoded amino acids sequences of the double-stranded oligonucleotide are as follows:

SEQ ID NO: 7

15 Bam HI

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5' GATCC GGT GGA GGT GGT TCT GGT GGA GGT GGT TCA GGT GGT GGA GGA TCA
3' G CCT CCT CCA CCA AGA CCA CCT CCA CCA AGT CCA CCA CCT CCT AGT
Ser Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser
  
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BspMII XbaI

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GGA GGT GGT GGA TCA GGT GGA GGA GGT TCT GGA GGT GGA GGT TCC GGA T 3'
25 CCT CCA CCA CCT AGT CCA CCT CCT CCA AGA CCT CCA CCT CCA AGG CCT AGATC
5'
Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser Gly
  
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30 The remaining portion of the linker is added during vector construction as described below. This oligonucleotide as well as those discussed below may be synthesized on an automated DNA synthesis machine such as those available from Biosearch, Inc., San Rafael, California or Applied Biosystems.

35 The linker encoding oligonucleotide is cloned into a vector that preferably contains multiple restriction endonuclease cleavage sites that may be employed for inserting the sequences encoding LIF-R and gp130 on either side of, and in the same reading frame as, the sequence encoding the linker. One such vector is designated pBLUESCRIPT SK<sup>®</sup> which is available from Stratagene Cloning Systems, La Jolla, California. This plasmid vector is replicable in *E. coli* and contains a polylinker segment that includes 21 unique restriction sites. The plasmid is digested with the  
 40 restriction enzymes BamHI and XbaI and the linker-encoding oligonucleotide is ligated into the vector using conventional techniques. A recombinant vector containing the inserted oligonucleotide sequence is identified by restriction endonuclease analysis and

sizing by gel electrophoresis. A DNA sequence encoding LIF-R is inserted into the pBLUESCRIPT SK<sup>®</sup> vector upstream of the linker- encoding oligonucleotide and a DNA sequence encoding gp130 is inserted downstream of the linker sequence. cDNA molecules encoding soluble fragments of LIF-R and gp130 were isolated and amplified using the well known polymerase chain reaction (PCR) procedure. The following oligonucleotides were synthesized for use in the PCR procedures:

SEQ ID NO: 8 (Oligonucleotide No. 1)

10                    SalI  
5'       GATATGTCGACGATGATGGATATTTACGTATGTTTG       3'

15 SEQ ID NO: 9 (Oligonucleotide No. 2)

3' CATACATACACCACTGTTTCCTTTTAAGACCTCCTCCACCTAGGTACG 5'  
BamHI

20

SEQ ID NO: 10 (Oligonucleotide No. 3)

25 5' BspMII CGCGTCCGGAGGAGGTGGATCTGAACCTCTAGATCCATGTGGTTATATC 3'

SEO ID No. 11      (Oligonucleotide No. 4)

3' CAAACGAGTTCCTCTTTAACTTATCCGCCGGCGTACG 5'

Oligonucleotides 1 and 2 are used in a PCR reaction to isolate a soluble fragment of LIF-R. The template employed in the reaction is the human LIF-R cDNA  
35 cloned as described by Gearing et al. *supra*. The DNA and encoded amino acid sequences of the cDNA clone are represented in SEQ ID NO: 5. The cloning vector which contains this human LIF-R cDNA clone was deposited in *E. coli* host cells with the American Type Culture Collection, Rockville, Maryland, U.S.A. on December 11, 1990, under the name pHLIFR-65 (ATCC Accession Number 68491). The deposit  
40 was made under the conditions of the Budapest Treaty. The 5' primer is oligonucleotide No. 1, which includes a DNA sequence encoding the first 8 amino acids of the signal sequence of LIF-R and also comprises upstream sequences that introduce a Sal I restriction endonuclease cleavage site. Oligonucleotide No. 1 is capable of annealing to the (-) strand that is complementary to nucleotides 179-202 of  
45 SEQ ID NO: 5. The 3' primer is oligonucleotide No. 2, which contains a sequence

complementary to nucleotides 2651-2677 of SEQ ID NO: 5 (i.e., includes anti-sense nucleotides encoding the last nine amino acids of the extracellular domain of LIF-R). Immediately downstream of the LIF-R encoding sequence, oligonucleotide No. 2 contains a sequence encoding (Gly)<sub>4</sub> Ser, and also introduces a BamHI restriction endonuclease cleavage site.

A PCR reaction employing oligonucleotides Nos. 1 and 2 thus isolates and amplifies a DNA sequence encoding an LIF-R fragment containing the entire signal sequence and the entire extracellular domain but lacking the transmembrane region and the extracellular domain. The (Gly)<sub>4</sub> Ser sequence attached to the 3' terminus of the LIF-R fragment is part of the polypeptide linker in the final construct.

Any suitable PCR procedure may be employed. One such procedure is described in Sarki et al., *Science* 239:487 (1988). Another is described in *Recombinant DNA Methodology*, Wu et al., eds., Academic Press Inc., San Diego (1989), pp. 189-196. In general, PCR reactions involve combining the 5' and 3' nucleotides with the template DNA and each of the four deoxynucleoside triphosphates in a suitable buffered solution. The solution is heated, (e.g. from 95° to 100°C) to denature the double-stranded DNA template and is then cooled before addition of a DNA polymerase enzyme. Multiple cycles of the reactions are carried out in order to amplify the desired DNA fragment.

An example of a suitable PCR procedure is as follows. All temperatures are in degrees centigrade. The following PCR reagents are added to a 1.5 ml Eppendorf microfuge tube: 10 µl of 10X PCR buffer (500 mM KCl, 100 mM Tris-HCl, pH 8.3 at 25°C, 25 mM MgCl<sub>2</sub>, and 1 mg/ml gelatin) (Perkin-Elmer Cetus, Norwalk, CN), 10 µl of a 2 mM solution containing each dNTP (2 mM dATP, 2 mM dCTP, 2 mM dGTP and 2 mM dTTP), 2.5 units (0.5 µl of standard 5000 units/ml solution) of *Taq* DNA polymerase (Perkin-Elmer Cetus), 50 ng of template DNA, 5 µl of a 20 µM solution of each of oligonucleotide primers 1 and 2, and 74.5 µl water to a final volume of 100 µl. The final mixture is then overlaid with 100 µl parafin oil. PCR is carried out using a DNA thermal cycler (Ericomp, San Diego, CA) by initially denaturing the template at 94° for 90 seconds, reannealing at 55° for 75 seconds and extending the cDNA at 72° for 150 seconds. PCR is carried out for an additional 20 cycles of amplification using a step program (denaturation at 94°, 25 sec; annealing at 55°, 45 sec; extension at 72°, 150 sec.), followed by a 5 minute extension at 72°.

The sample is removed from the parafin oil and DNA extracted by phenolchloroform extraction and spun column chromatography over G-50 (Boehringer Mannheim). A 10 µl aliquot of the extracted DNA is separated by electrophoresis on

1% SeaKem™ agarose (FMC BioProducts, Rockland, ME) and stained with ethidium bromide to confirm that the DNA fragment size is consistent with the predicted product.

The PCR-amplified cDNA products are then digested with SalI and BamHI restriction enzymes using standard procedures. The SalI/BamHI restriction fragment is then separated by gel electrophoresis, e.g., on a 1.2% Seaplaque™ low gelling temperature (LGT) agarose, and the band representing the desired fragment is isolated. The fragment is inserted into a vector encoding the desired fusion protein as described below.

A plasmid vector containing human gp130 cDNA was deposited in *E. coli* strain DH5α host cells with the American Type Culture Collection, Rockville, Maryland under the name B10G/pDC303 (DH5α) on November 14, 1991 and assigned ATCC Accession No. 68827. The deposit was made under the conditions of the Budapest Treaty. The DNA and encoded amino acid sequences of this cloned cDNA are shown in SEQ ID NO: 1.

Oligonucleotides 3 and 4 are employed in the polymerase chain reaction procedure to amplify and isolate a DNA fragment encoding Ser(Gly)<sub>4</sub> Ser followed by amino acids 1 to 597 of SEQ ID NO: 1 (the entire extracellular domain of the mature gp130 protein). The 5' primer, oligonucleotide No. 3, includes nucleotides 310 to 336 of SEQ ID NO: 1, which encode the first nine amino acids of the mature gp130 protein. This nucleotide sequence is capable of annealing to the (-) strand that is complementary to nucleotides 310 to 336 of SEQ ID NO: 1. Oligonucleotide No. 3 also encodes a Ser(Gly)<sub>4</sub> Ser sequence directly upstream of (and in the same reading frame as) the gp130 sequence, and further positions a BspMII restriction endonuclease cleavage site near the 5' terminus of the Ser(Gly)<sub>4</sub> Ser -encoding sequence.

The 3' primer, oligonucleotide No. 4, includes a sequence complimentary to nucleotides 2080 to 2100 of SEQ ID NO: 1, i.e., includes anti-sense nucleotides encoding the last seven amino acids of the gp130 extracellular domain. Oligonucleotide No. 4 positions a stop codon immediately after the gp130 sequence and also inserts a NotI restriction site downstream. Following amplification of the gp130 fragment by PCR, the PCR reaction products are digested with BspMII and NotI and the desired fragment is isolated.

The above-described LIF-R, Ser(Gly<sub>4</sub>Ser)<sub>6</sub>Gly linker, and gp130 encoding fragments are assembled into a single DNA sequence as follows. The Ser(Gly<sub>4</sub>Ser)<sub>6</sub>Gly linker fragment is excised from the pBLUESCRIPT SK® vector by digestion with BamHI and BspMII. The linker fragment is then ligated to the 3' end of the LIF-R fragment (cleaved at its 3' terminus after the Gly<sub>4</sub>Ser sequence with BamHI). The ligation is conducted under conventional conditions. The 3' end of the

linker fragment is ligated to the BspMII-cleaved 5' end of the gp130 fragment. The resulting DNA fragment encodes a receptor of the present invention comprising (from 5' to 3') the signal sequence and extracellular domain of LIF-R attached to a (Gly<sub>4</sub>Ser)<sub>8</sub> polypeptide linker which is attached to the mature coding sequence of the gp130 extracellular domain.

5 This DNA fragment may be inserted into any suitable cloning and/or expression vector. For example, the pBLUESCRIPT SK<sup>®</sup> vector may be digested with SalI and NotI and the ligated DNA fragment inserted therein. *E. coli* cells are then transformed with the recombinant vector by conventional procedures.

10 In an alternative procedure, the pBLUESCRIPT SK<sup>®</sup> vector containing the Ser(Gly<sub>4</sub>Ser)<sub>6</sub>Gly linker sequence is digested with SalI and BamHI and the above described LIF-R-encoding fragment is inserted therein. The resulting vector is then digested with BspMII and NotI and the gp130-encoding fragment is then inserted to form the DNA sequence encoding the receptor of the present invention. The cloned  
15 receptor-encoding DNA fragment may be excised and inserted into any suitable expression vector (chosen according to the type of host cell that is desired) using conventional procedures. Host cells transformed with the recombinant expression vector are cultivated to produce the receptor protein. Mammalian host cells are generally preferred for producing the recombinant receptor fusion proteins of the  
20 present invention.

The receptor-encoding construct may be excised by SalI and NotI digestion and inserted into a vector suitable for use in mammalian host cells. One suitable vector is designated pDC406. cDNA molecules inserted at the SalI site of this vector are transcribed and translated using regulatory elements derived from HIV and adenovirus.  
25 pDC406 contains origins of replication derived from SV40, Epstein-Barr virus and pBR322. The pDC406 vector into which interleukin-1 receptor cDNA has been cloned has been deposited with the American Type Culture Collection, Rockville, Maryland USA under accession number CRL10478. The interleukin-1 receptor cDNA may be excised from the vector using conventional techniques and replaced with the receptor  
30 encoding DNA of the present invention prepared above. pDC406 is a derivative of HAV-EO described by Dower et al., *J. Immunol.* 142:4314 (1989). pDC406 differs from HAV-EO by the deletion of the intron present in the adenovirus 2 tripartite leader sequence in HAV-EO.

35 Examples of suitable mammalian cells for expressing a receptor fusion protein include CV-1 cells (ATCC CCL70) and COS-7 cells, (ATCC CRL 1651) both derived from monkey kidney. Another monkey kidney cell line CV-1/EBNA (ATCC CRL 10478) was derived by transfection of the CV-1 cell line with a gene encoding Epstein-

Barr virus nuclear antigen-1 (EBNA-1) and with a vector containing CMV regulatory sequences. See McMahan et al., *EMBO J.* 10:2821 (1991). The EBNA-1 gene allows for episomal replication of expression vectors, such as HAV-EO or pDC406, that contain the EBV origin of replication.

5

#### Example 4

##### Preparation of a Recombinant Receptor Fusion Protein Designating gp130-Linker-LIF-R

10 This receptor of the present invention differs from that of Example 3 in that the LIF-R polypeptide (which was the 5' polypeptide in the receptor of Example 3) is now the 3' polypeptide. The following oligonucleotides were synthesized for use in preparing the fusion protein:

15 SEQ ID NO: 12

5' GATATGTCGACAAGATGTTGACGTTGCAGACTTGG 3' (oligonucleotide no. 5)

SEQ ID NO: 13

3' CAAACGAGTTCCTCTTTAACTTCCTCCTCCACCTAGGTACG 5' (oligonucleotide no. 6)

20

SEQ ID NO: 14

5' CGCGTCCGGAGGAGGTGGTAGCCAGAAAAAGGGGCTCCTCATG 3' (oligonucleotide no. 7)

25 SEQ ID NO: 15

3' CATACATACCACTGTTTCCTTTTAAGAATCGCCGGCGTACG 5' (oligonucleotide no. 8)

30 Oligonucleotides 5 and 6 are employed in a polymerase chain reaction procedure to isolate a fragment of gp130. The 5' primer (oligonucleotide number 5) includes nucleotides 244 to 264 of SEQ ID NO: 1, (the sequence encoding the first seven amino acids of the gp130 signal sequence). Oligonucleotide number 5 also includes a sequence that introduces an upstream SalI site. This nucleotide sequence is capable of annealing to the (-) strand that is complementary to nucleotides 244 to 264 of SEQ ID  
35 NO: 1. The 3' primer (oligonucleotide number 6) includes a sequence complementary to nucleotides 2080 to 2100 of SEQ ID NO: 1, i.e. includes antisense nucleotides encoding the last seven amino acids of the gp-130 extracellular domain.

Oligonucleotide number 6 also encodes a Gly<sub>4</sub>Ser sequence immediately 3' to (and in phase with) the gp130 sequence, and also inserts a downstream BamHI site.

5 A PCR reaction is conducted as described in Example 3 but employing oligonucleotides 5 and 6 on the gp130 cDNA template. A DNA sequence encoding a gp130 fragment that includes the 5' signal sequence and the entire extracellular domain, but none of the transmembrane region or the cytoplasmic domain, is isolated by the PCR reaction. A Gly<sub>4</sub>Ser sequence is fused to the 3' terminus of the gp130 fragment. The PCR reaction products are digested with Sall and BamHI and the desired fragment is isolated.

10 An LIF-R fragment is isolated and amplified by a PCR reaction employing oligonucleotides 7 and 8. The 5' primer (oligonucleotide number 7) includes nucleotides 311 to 331 of SEQ ID NO: 5, which encode the first seven amino acids of the mature LIF-R protein. This nucleotide sequence is capable of annealing to the (-) strand complementary to nucleotides 311 to 331 of SEQ ID NO: 5. Oligonucleotide  
15 number 7 also encodes a Gly<sub>4</sub>Ser sequence fused to the 5' end of the LIF-R sequence, and inserts an upstream BspMII site. The 3' primer (oligonucleotide number 8) is complementary to nucleotides 2651 to 2677 of SEQ ID NO: 5 (which encode the last nine amino acids of the LIF-R extracellular domain.) Oligonucleotide number 8 also adds a stop codon at the 3' end of the LIF-R sequence, and inserts a Not I site  
20 downstream. The PCR reaction products are digested with BspMII and NotI and the desired fragment is isolated.

A DNA sequence encoding the desired receptor protein is prepared by ligating the BamHI site of the gp130 fragment prepared above to the BamHI site at the 5' terminus of the linker fragment described in Example 3. Likewise the C-terminus of  
25 the linker encoding fragment is ligated at the BspMII site to the complementary site of the LIF-R encoding fragment prepared above. The resulting DNA fragment may be cloned into an expression vector using procedures described in Example 3. The receptor encoded by the isolated DNA fragment comprises (from the N-terminus to the C-terminus) the signal sequence and extracellular domain of gp130 attached to a  
30 (Gly<sub>4</sub>Ser)<sub>8</sub> polypeptide linker which is attached to the mature coding sequence of the LIF-R extracellular domain.

**Example 5****Receptor Fusion Protein Comprising LIF-R Attached to gp-130 Through An Fc Polypeptide Linker**

5

A receptor prepared in accordance with the following procedures is depicted in Figure 4. The following oligonucleotides were synthesized for use in preparing the receptor fusion protein:

SEQ ID NO: 16

10        3' CATACATACACCACTGTTTCCTTTAAGACTCGGGTCTAGATACG 5' (oligonucleotide no. 9)

SEQ ID NO: 17

3' CAAACGAGTTCCTCTTTAACTTCTCGGGTCTAGATACG 5' (oligonucleotide no. 10)

15

An LIF-R encoding DNA sequence is isolated and amplified in a PCR reaction using oligonucleotides 1 and 9. Oligonucleotide number 1 (the 5' primer) inserts an upstream SalI site and has been described in Example 3. The 3' primer is oligonucleotide number 9 which includes a sequence complementary to nucleotides  
20        2651 to 2677 of SEQ ID NO: 5, i.e., includes antisense nucleotides encoding the last nine amino acids of the extracellular domain of LIF-R. Oligonucleotide number 9 also inserts a downstream BglII site. The PCR reaction products are digested with SalI and BglII, and the desired LIF-R encoding DNA fragment is isolated by gel electrophoresis using conventional procedures. Due to the presence of an internal BglII site in the LIF-R sequence, the BglII digestion should be carried out under conditions that effect partial  
25        digestion.

A gp130 encoding DNA fragment is isolated and amplified by PCR reaction using oligonucleotides 5 and 10. The 5' primer (oligonucleotide number 5) inserts an upstream SalI site and has been described above in Example 4. The 3' primer is  
30        oligonucleotide number 10, which includes a sequence complementary to nucleotides 2080 to 2100 of SEQ ID NO: 1, i.e., includes antisense nucleotides encoding the last seven amino acids of the gp130 extracellular domain. Oligonucleotide number 10 also inserts a downstream BglII site. The PCR reaction products are digested with SalI and BglII, and the desired gp130 encoding DNA fragment is isolated by gel electrophoresis  
35        using conventional techniques.

cDNA encoding a single chain polypeptide derived from the Fc region of a human IgG1 antibody has been cloned into the above-described pBLUESCRIPT SK®

vector to produce a recombinant vector designated hIgG1Fc. A unique BglII site is positioned near the 5' end of the inserted Fc encoding sequence. An SpeI site is immediately downstream of the stop codon. The DNA and encoded amino acid sequences of the cloned Fc cDNA are presented in SEQ ID NO: 3 and SEQ ID NO: 4.

5       The Fc polypeptide encoded by the cDNA extends from the N-terminal hinge region to the native C-terminus, i.e., is an essentially full-length antibody Fc region. Fc fragments, e.g., those that are truncated at the C-terminal end, also may be employed. The fragments should contain multiple cysteine residues (at least the cysteine residues in the hinge region). The antibody from which the Fc polypeptide is  
10       derived is preferably of the same species as the patient to be treated with the fusion protein prepared therefrom..

Plasmid hIgG1Fc is digested with BglII and SalI and the BglII/SalI LIF-R fragment prepared above is ligated into the vector by conventional techniques. The Fc encoding sequence is positioned downstream of, and in the same reading frame as, the  
15       LIF-R sequence. In a separate reaction, the above-described SalI/BglII fragment of gp130 is also inserted into the same vector. Plasmid vectors containing the desired DNA insert are identified by restriction endonuclease digestion analysis, using convention techniques.

The cloned DNA segment encoding the LIF-R-Fc fusion polypeptide may be  
20       excised from the pBLUESCRIPT SK<sup>®</sup> vector by digestion with SalI and NotI. Likewise, the DNA segment encoding the gp130-Fc fusion polypeptide may be excised by SalI/NotI digestion. Each excised DNA segment is inserted into an appropriate expression vector, depending on the type of host cell that is desired. One suitable expression vector is the plasmid pDC406, which may be transformed into mammalian  
25       host cells as described in Example 3.

In one embodiment of the invention, an expression vector encoding the LIF-R-Fc fusion and a second expression vector encoding the gp130-Fc fusion are co-transfected into the desired host cells. Two separate recombinant polypeptides are thus produced in the host cells. The first polypeptide comprises the Fc polypeptide fused in  
30       frame to the C-terminus of the gp130 fragment. The second polypeptide comprises the Fc polypeptide fused in frame to the C-terminus of the LIF-R fragment. Disulfide bonds that form between the two Fc regions covalently link the two separate fusion polypeptides into a receptor protein of the present invention.

Alternatively, the LIF-R-Fc and gp130-Fc polypeptides may be separately  
35       transformed into host cells (as opposed to co-transfection into the same host cell.) The two polypeptides are purified from the host cells and then combined in a suitable

buffered solution, whereupon interchain disulfide bonds form between the two Fc regions.

The receptor protein may be purified using any of a number of conventional protein purification techniques. Since antibody Fc regions bind to protein A and protein G, affinity chromatography employing protein A or protein G attached to an insoluble support material may be employed in the purification process. In one procedure, one liter of culture supernatant containing the receptor is passed over a solid phase protein G column, and the column is then washed thoroughly with phosphate-buffered saline (PBS). The adsorbed Fc-containing fusion protein is eluted with 50 mM glycine buffer, pH 3 and brought to pH 7 with 2 M Tris buffer, pH 9. Further purification may involve immunoaffinity column(s), e.g., affinity columns having LIF or OSM bound thereto.

#### Example 6

##### Preparation of Monoclonal Antibodies Directed against a Receptor

Preparations of a purified receptor protein of the present invention, or transfected COS cells expressing high levels of the receptor, are employed to generate monoclonal antibodies against the receptor using conventional techniques, for example, those disclosed in U.S. Patent 4,411,993. To immunize mice, a receptor immunogen is emulsified in complete Freund's adjuvant and injected subcutaneously in amounts ranging from 10-100 $\mu$ g into Balb/c mice. Ten to twelve days later, the immunized animals are boosted with additional immunogen emulsified in incomplete Freund's adjuvant and periodically boosted thereafter on a weekly to biweekly immunization schedule. Serum samples are periodically taken by retro-orbital bleeding or tail-tip excision for testing by dot-blot assay (antibody sandwich) or ELISA (enzyme-linked immunosorbent assay). Other assay procedures are also suitable. Following detection of an appropriate antibody titer, positive animals are given an intravenous injection of antigen in saline. Three to four days later, the animals are sacrificed, splenocytes harvested, and fused to the murine myeloma cell line NS1. Hybridoma cell lines generated by this procedure are plated in multiple microtiter plates in a HAT selective medium (hypoxanthine, aminopterin, and thymidine) to inhibit proliferation of non-fused cells, myeloma hybrids, and spleen cell hybrids.

Hybridoma clones thus generated can be screened by ELISA for reactivity with the receptor protein, for example, by adaptations of the techniques disclosed by Engvall et al., *Immunochem* 8.871 (1971) and in U.S. Patent 4,704,004. Positive clones are then injected into the peritoneal cavities of syngeneic Balb/c mice to produce ascites

containing high concentrations (greater than 1 mg/ml) of anti-receptor monoclonal antibody. The resulting monoclonal antibody can be purified by ammonium sulfate precipitation followed by gel exclusion chromatography, and/or affinity chromatography based on binding of antibody to Protein A of *Staphylococcus aureus*.

5

### Example 7

#### Heterodimeric Receptor

An expression vector encoding a fragment of the human LIF-R extracellular domain fused to a polypeptide derived from the Fc region of an antibody was constructed as follows. A second expression vector encoding a fragment of the human gp130 extracellular domain fused to an Fc polypeptide also was constructed.

Plasmid pHLIF-R-65 (ATCC 68491), which contains human LIF-R cDNA in expression vector pDC303 as described in example 3, was digested with the restriction enzymes *Asp718* and *XmnI*. *Asp718* cleaves the vector upstream of the LIF-R cDNA insert. *XmnI* is a blunt cutter that cleaves within the codon for amino acid number 702 (Asp) of SEQ ID NO:5, upstream of the transmembrane region. The desired *Asp718/XmnI* fragment (about 2,444 bp in length) was separated by electrophoresis on an agarose gel and purified by conventional procedures using an Elutip column.

A recombinant vector designated hIgG1Fc, comprising cDNA encoding a single chain polypeptide derived from the Fc region of a human IgG1 antibody in a pBLUESCRIPT SK® vector was described in example 5. The DNA and encoded amino acid sequences of the cloned Fc cDNA are presented in SEQ ID NO:3 and SEQ ID NO:4. A polylinker region comprising a number of restriction sites is positioned immediately upstream of the Fc cDNA.

Plasmid hIgG1Fc was digested with *Asp718* and *StuI*, which cleave within the polylinker upstream of the Fc sequence. The *Asp718/XmnI* LIF-R fragment prepared above was ligated into the cleaved hIgG1Fc vector by conventional techniques. *StuI* and *XmnI* both produce blunt ends, which will ligate together. In the resulting recombinant vector, the Fc encoding sequence is positioned downstream of, and in the same reading frame as, the LIF-R sequence. The encoded LIF-R/Fc fusion protein comprises amino acids -44 to 702 of SEQ ID NO:5 (LIF-R), followed by six amino acids constituting a peptide linker encoded by the polylinker segment of plasmid hIgG1Fc, followed by amino acids 1-232 of SEQ ID NO:3 (Fc). *E. coli* cells were transformed with the ligation mixture and plasmids were isolated therefrom by standard procedures. Plasmid vectors containing the desired DNA insert were identified by restriction endonuclease digestion analysis.

The cloned DNA segment encoding the LIF-R/Fc fusion polypeptide was excised from the recombinant vector by digestion with *Asp718* and *NotI*. The *NotI* enzyme cleaves the vector in a polylinker region just downstream of the Fc cDNA insert. The excised DNA segment (3.2 kb) is inserted into an appropriate expression  
5 vector, depending on the type of host cell that is desired. One suitable expression vector is pCAV/NOT, a mammalian expression vector described in PCT application WO 90/05183.

pCAV/NOT was cleaved with *Asp718* and *NotI*, both of which cleave in the multiple cloning site. The LIF-R/Fc-encoding *Asp718/NotI* DNA fragment prepared  
10 above was ligated into the vector.

An expression vector encoding a soluble gp130/Fc fusion protein was constructed as follows. Recombinant vector B10G/pDC303 (ATCC 68827) comprising human gp130 cDNA (described in example 3) was digested with *EcoR1*, and the resulting 5' overhang was rendered blunt using T4 DNA polymerase. The  
15 recognition site for *EcoR1* comprises nucleotides 2056-2061 of SEQ ID NO:1. The *EcoR1*-digested vector was then cleaved with *XhoI*, which cleaves in the vector upstream of the gp130 cDNA insert.

Plasmid hIgG1Fc, comprising Fc polypeptide-encoding cDNA as described above, was digested with *StuI* (a blunt cutter) and *NotI*, which cleave upstream and  
20 downstream, respectively, of the inserted Fc cDNA. The (*EcoR1*)/*XhoI* gp130 fragment isolated above was ligated to the Fc-containing fragment and to *XhoI/NotI*-digested vector SF CAV/NOT. The mammalian expression vector SF CAV/NOT is essentially identical to SF CAV (ATCC 68922) but contains a *NotI* site. SF CAV/NOT also is essentially identical to pCAV/NOT, described in PCT application WO 90/05183,  
25 except that a segment of the adenovirus-2 tripartite leader (TPL) containing a cryptic promoter functional in bacteria has been deleted. Protein expression from the cryptic promoter is potentially disadvantageous for preparing and isolating a desired recombinant plasmid in bacterial cells.

*E. coli* cells were transformed with the ligation mixture, plasmids were isolated  
30 therefrom by conventional procedures, and the desired recombinant plasmids were identified by restriction analysis. The gp130/Fc fusion protein encoded by the desired recombinant vector comprises (from N- to C-terminus) amino acids -22 to 582 of SEQ ID NO:2 (gp130), followed by 7 amino acids constituting a peptide linker encoded by the polylinker segment of plasmid hIgG1Fc, followed by amino acids 1-232 of SEQ ID  
35 NO:4 (Fc).

COS-7 cells (ATCC CRL 1651) were transfected with either the LIF-R/Fc-encoding recombinant expression vector or the gp130/Fc-encoding expression vector

prepared above, or with both expression vectors. The cells were cultivated to allow expression of the soluble fusion proteins. The expressed proteins were recovered by incubating culture supernatant with Protein G Sepharose beads (available from Pharmacia) overnight at 4°C, then pelleting the beads by centrifugation. The binding of <sup>125</sup>I-labeled human oncostatin M and <sup>125</sup>I-labeled human LIF by the proteins bound to the beads was analyzed.

Binding affinity was determined by performing a variation of a standard Scatchard analysis. The binding assay procedure was similar to that described by Mosley et al. (*Cell* 59:335, 1989) except that the proteins, being soluble, are attached to Protein G Sepharose beads rather than being on the surface of the transfected cells. Briefly, in a 96-well microtiter plate, each of ten 1:2 serial dilutions of <sup>125</sup>I-LIF or <sup>125</sup>I-oncostatin M was incubated with a sample comprising the expressed proteins (bound to the beads) resuspended in RPMI 1640 containing 2.5% bovine serum albumin, 0.2% (v/v) sodium azide and 20mM Hepes, pH 7.4, for 2 hours at 4°C with agitation. Duplicate standard cold competition wells also were incubated. Centrifuge tubes containing bovine calf serum were used in place of the phthalate oil mixture-containing tubes in the separation method described by Dower et al., *J. Immunol.* 132:751 (1984) and Park et al., *J. Biol. Chem.* 261:4177 (1986) and in example 1 above. Aliquots of each incubation mixture were transferred to the tubes. After centrifugation, tubes were cut, the radioactivity counted, and processed as for standard Scatchard analysis.

Figure 7 presents Scatchard analyses of the binding of <sup>125</sup>I oncostatin M by gp130/Fc homodimers produced by the cells transfected with the gp130/Fc vector alone (upper left) and by the proteins expressed by the co-transfected cells (lower left). Scatchard analyses of the binding of <sup>125</sup>I LIF by LIF-R/Fc homodimers produced by the cells transfected with the LIF-R/Fc vector alone (upper right) and by the proteins expressed by the co-transfected cells (lower right) are also presented in figure 7. A shift toward higher affinity binding of oncostatin M by the proteins recovered from the co-transfected cells, compared to the gp130/Fc homodimer, is evident from figure 7. Likewise, the data in figure 7 indicate a shift toward higher affinity binding of LIF by the proteins recovered from the co-transfected cells, compared to the LIF-R/Fc homodimer. The shift toward higher affinity binding indicates the presence of heterodimers comprising LIF-R/Fc and gp130/Fc, and further indicates that the LIF-R and gp130 moieties are cooperating, i.e., interacting, in the binding of oncostatin M and LIF. Controls demonstrated no oncostatin M binding by LIF-R homodimers, and no LIF binding by gp130 homodimers.

**Example 8****Receptors Comprising LIF-R and gp130 Polypeptides Lacking FNIII Domains**

DNA sequences encoding soluble LIF-R and gp130 proteins lacking fibronectin type III (FNIII) domains were isolated and fused to an Fc-encoding sequence. Deleting the FNIII domains affords the advantage of reducing the size of the LIF-R/Fc and gp130/Fc fusion proteins. The LIF-R protein of SEQ ID NO:6 comprises three repeats of a fibronectin type III-like module in the extracellular domain. The three domains containing FNIII modules comprise amino acids 487 (Thr) to 584 (Asn), 585 (Asp) to 679 (Ala), and 680 (Pro) to 789 (Ser), respectively, of SEQ ID NO:6. gp130 also contains three FNIII domains, comprising amino acids 300 (Tyr) to 399 (Phe), 400 (Gln) to 496 (Pro), and 497 (Pro) to 597 (Glu), respectively, of SEQ ID NO:2. From one to all three of the FNIII domains may be removed from gp130 or LIF-R to reduce the size of the protein.

The FNIII domains of human LIF-R were removed by digesting the LIF-R/Fc-encoding expression vector prepared in example 7 with the restriction endonuclease Eco O 109I (isoschizomer of Dra II) and filling in the resulting overhangs using T4 DNA polymerase according to conventional procedures. The recognition site for Eco O 109I spans nucleotides 1789-1795 of SEQ ID NO:5 (LIF-R), cleaving within the codons for amino acids 8-9 of the first FNIII domain of LIF-R. The cleaved vector was then digested with BstX1 and EcoR5. The recognition site for BstX1 spans nucleotides 1048-1059 of SEQ ID NO:5 and EcoR5 (which generates blunt ends) cleaves within the polylinker upstream of the Fc sequence. The BstX1/EcoR5 fragment (comprising the 5' end of LIF-R, the vector, the entire Fc sequence, and a portion of the polylinker) and the BstX1/(Eco O 109I) LIF-R fragment were isolated and ligated together. *E. coli* cells were transformed with the ligation mixture, plasmids were isolated therefrom, and the desired recombinant plasmid was identified by restriction analysis. The resulting construct encodes a fusion protein comprising (from N- to C-terminus) amino acids -44 to 494 of SEQ ID NO:5 (LIF-R), a four amino acid spacer peptide -His-Arg-Tyr-Val- encoded by the polylinker segment, and amino acids 1-232 of SEQ ID NO:3 (Fc). The LIF-R polypeptide moiety contains the first 8 amino acids of the first FNIII domain, but lacks the remainder of the first FNIII domain and all of the second and third FNIII domains.

The FNIII domains of gp130 were removed by digesting the recombinant gp130/Fc-encoding expression vector prepared in example 7 with BstX1, then blunting the overhang using T4 DNA polymerase according to conventional procedures. The recognition site for BstX1 spans nucleotides 1231-1242 of SEQ ID NO:1 (gp130),

cleaving within the codons for amino acids 10-11 of the first FNIII domain of gp130. The cleaved vector was then digested with EcoR5, which cleaves within the polylinker upstream of the Fc sequence and generates blunt ends. The (BstX1)/EcoR5 fragment comprising the 5' end of gp130 (lacking the FNIII domains), the vector sequences, the Fc sequence, and a portion of the polylinker, was ligated. *E. coli* cells were transformed with the ligation mixture, plasmids were isolated therefrom, and the desired recombinant plasmid was identified by restriction analysis. The fusion protein encoded by the construct comprises (from N- to C-terminus) amino acids -22 to 308 of SEQ ID NO:2 (gp130), a four amino acid spacer peptide -Asn-Arg-Tyr-Val- encoded by the polylinker segment, and amino acids 1-232 of SEQ ID NO:3 (Fc). The gp130 polypeptide moiety contains the first 9 amino acids of the first FNIII domain, but lacks the remainder of the first FNIII domain and all of the second and third FNIII domains.

FNIII domains may be deleted from the gp130 component of a receptor of the present invention, from the LIF-R component, or from both. In one embodiment of the invention, COS-7 cells were co-transfected with the soluble LIF-R/Fc-encoding mammalian expression vector prepared in example 7 and the mammalian expression vector encoding a soluble gp130/Fc protein lacking the FNIII domains prepared above. Analysis of the expressed proteins by SDS-PAGE revealed a band of the molecular weight expected for the heterodimer, along with bands that include those of the molecular weight expected for the two homodimers. Scatchard analyses conducted according to the procedures described in example 7 demonstrated a shift toward higher affinity binding of LIF and oncostatin M for proteins expressed by the co-transfected cells compared to the corresponding homodimers. This result indicates the presence of heterodimers comprising LIF-R/Fc and gp130/Fc, and further indicates that the LIF-R and gp130 moieties are cooperating, i.e., interacting, in binding oncostatin M and LIF.

### **BRIEF DESCRIPTION OF THE SEQUENCE LISTING**

SEQ ID NO:1 and SEQ ID NO:2 present the DNA sequence and encoded amino acid sequence for cloned cDNA encoding an N-terminal fragment of gp130.

SEQ ID NO:3 and SEQ ID NO:4 present the DNA sequence and encoded amino acid sequence for cloned cDNA encoding a polypeptide that corresponds to the Fc region of an IgG1 antibody.

SEQ ID NO:5 and SEQ ID NO:6 present the DNA sequence and encoded amino acid sequence for cloned cDNA encoding an N-terminal fragment of LIF-R.

SEQ ID NO:7 presents the DNA sequence of the coding strand of a chemically synthesized DNA molecule encoding a polypeptide linker used in constructing certain receptors of the present invention.

5

SEQ ID NO:8 - SEQ ID NO:17 present the DNA sequence of various single-stranded oligonucleotide primers employed in polymerase chain reactions to construct certain receptors of the present invention.

## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

- (i) APPLICANT: Gearing, David P.
- (ii) TITLE OF INVENTION: Receptor for Oncostatin M and Leukemia Inhibitory Factor
- (iii) NUMBER OF SEQUENCES: 17
- (iv) CORRESPONDENCE ADDRESS:
  - (A) ADDRESSEE: Immunex Corporation
  - (B) STREET: 51 University Street
  - (C) CITY: Seattle
  - (D) STATE: WA
  - (E) COUNTRY: USA
  - (F) ZIP: 98101
- (v) COMPUTER READABLE FORM:
  - (A) MEDIUM TYPE: Floppy disk
  - (B) COMPUTER: IBM PC compatible
  - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
  - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
  - (A) APPLICATION NUMBER:
  - (B) FILING DATE:
  - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
  - (A) NAME: Seese, Kathryn A.
  - (B) REGISTRATION NUMBER: 32,172
  - (C) REFERENCE/DOCKET NUMBER: 2607
- (ix) TELECOMMUNICATION INFORMATION:
  - (A) TELEPHONE: 206-587-0430
  - (B) TELEFAX: 206-587-0606

## (2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2369 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (v) FRAGMENT TYPE: N-terminal
- (vi) ORIGINAL SOURCE:
  - (F) TISSUE TYPE: human placenta
- (vii) IMMEDIATE SOURCE:

48

(B) CLONE: B10G/pDC303

## (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 244..2369

## (ix) FEATURE:

(A) NAME/KEY: mat\_peptide

(B) LOCATION: 310..2369

## (ix) FEATURE:

(A) NAME/KEY: sig\_peptide

(B) LOCATION: 244..309

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

```

GGCCCGCGGA GTCGCGCTGG GCCGCCCCGG CGCAGCTGAA CCGGGGGCCG CGCCTGCCAG      60
GCCGACGGGT CTGGCCAGC CTGGCGCCAA GGGGTTTCGTG CGCTGTGGAG ACGCGGAGGG      120
TCGAGGCGGC GCGGCCTGAG TGAAACCCAA TGGAAAAAGC ATGACATTTA GAAGTAGAAG      180
ACTTAGCTTC AAATCCCTAC TCCTTCACTT ACTAATTTTG TGATTGGAA ATATCCGCGC      240
AAG ATG TTG ACG TTG CAG ACT TGG CTA GTG CAA GCC TTG TTT ATT TTC      288
  Met Leu Thr Leu Gln Thr Trp Leu Val Gln Ala Leu Phe Ile Phe
    -22      -20                      -15                -10

CTC ACC ACT GAA TCT ACA GGT GAA CTT CTA GAT CCA TGT GGT TAT ATC      336
Leu Thr Thr Glu Ser Thr Gly Glu Leu Leu Asp Pro Cys Gly Tyr Ile
      -5                      1                5

AGT CCT GAA TCT CCA GTT GTA CAA CTT CAT TCT AAT TTC ACT GCA GTT      384
Ser Pro Glu Ser Pro Val Val Gln Leu His Ser Asn Phe Thr Ala Val
  10                15                20                25

TGT GTG CTA AAG GAA AAA TGT ATG GAT TAT TTT CAT GTA AAT GCT AAT      432
Cys Val Leu Lys Glu Lys Cys Met Asp Tyr Phe His Val Asn Ala Asn
          30                35                40

TAC ATT GTC TGG AAA ACA AAC CAT TTT ACT ATT CCT AAG GAG CAA TAT      480
Tyr Ile Val Trp Lys Thr Asn His Phe Thr Ile Pro Lys Glu Gln Tyr
          45                50                55

ACT ATC ATA AAC AGA ACA GCA TCC AGT GTC ACC TTT ACA GAT ATA GCT      528
Thr Ile Ile Asn Arg Thr Ala Ser Ser Val Thr Phe Thr Asp Ile Ala
      60                65                70

TCA TTA AAT ATT CAG CTC ACT TGC AAC ATT CTT ACA TTC GGA CAG CTT      576
Ser Leu Asn Ile Gln Leu Thr Cys Asn Ile Leu Thr Phe Gly Gln Leu
      75                80                85

GAA CAG AAT GTT TAT GGA ATC ACA ATA ATT TCA GGC TTG CCT CCA GAA      624
Glu Gln Asn Val Tyr Gly Ile Thr Ile Ile Ser Gly Leu Pro Pro Glu
      90                95                100                105

AAA CCT AAA AAT TTG AGT TGC ATT GTG AAC GAG GGG AAG AAA ATG AGG      672
Lys Pro Lys Asn Leu Ser Cys Ile Val Asn Glu Gly Lys Lys Met Arg
          110                115                120

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49

TGT	GAG	TGG	GAT	GGT	GGA	AGG	GAA	ACA	CAC	TTG	GAG	ACA	AAC	TTC	ACT	720
Cys	Glu	Trp	Asp	Gly	Gly	Arg	Glu	Thr	His	Leu	Glu	Thr	Asn	Phe	Thr	
			125					130					135			
TTA	AAA	TCT	GAA	TGG	GCA	ACA	CAC	AAG	TTT	GCT	GAT	TGC	AAA	GCA	AAA	768
Leu	Lys	Ser	Glu	Trp	Ala	Thr	His	Lys	Phe	Ala	Asp	Cys	Lys	Ala	Lys	
			140					145				150				
CGT	GAC	ACC	CCC	ACC	TCA	TGC	ACT	GTT	GAT	TAT	TCT	ACT	GTG	TAT	TTT	816
Arg	Asp	Thr	Pro	Thr	Ser	Cys	Thr	Val	Asp	Tyr	Ser	Thr	Val	Tyr	Phe	
			155				160					165				
GTC	AAC	ATT	GAA	GTC	TGG	GTA	GAA	GCA	GAG	AAT	GCC	CTT	GGG	AAG	GTT	864
Val	Asn	Ile	Glu	Val	Trp	Val	Glu	Ala	Glu	Asn	Ala	Leu	Gly	Lys	Val	
			170				175				180				185	
ACA	TCA	GAT	CAT	ATC	AAT	TTT	GAT	CCT	GTA	TAT	AAA	GTG	AAG	CCC	AAT	912
Thr	Ser	Asp	His	Ile	Asn	Phe	Asp	Pro	Val	Tyr	Lys	Val	Lys	Pro	Asn	
				190					195					200		
CCG	CCA	CAT	AAT	TTA	TCA	GTG	ATC	AAC	TCA	GAG	GAA	CTG	TCT	AGT	ATC	960
Pro	Pro	His	Asn	Leu	Ser	Val	Ile	Asn	Ser	Glu	Glu	Leu	Ser	Ser	Ile	
			205					210					215			
TTA	AAA	TTG	ACA	TGG	ACC	AAC	CCA	AGT	ATT	AAG	AGT	GTT	ATA	ATA	CTA	1008
Leu	Lys	Leu	Thr	Trp	Thr	Asn	Pro	Ser	Ile	Lys	Ser	Val	Ile	Ile	Leu	
			220				225					230				
AAA	TAT	AAC	ATT	CAA	TAT	AGG	ACC	AAA	GAT	GCC	TCA	ACT	TGG	AGC	CAG	1056
Lys	Tyr	Asn	Ile	Gln	Tyr	Arg	Thr	Lys	Asp	Ala	Ser	Thr	Trp	Ser	Gln	
			235				240				245					
ATT	CCT	CCT	GAA	GAC	ACA	GCA	TCC	ACC	CGA	TCT	TCA	TTC	ACT	GTC	CAA	1104
Ile	Pro	Pro	Glu	Asp	Thr	Ala	Ser	Thr	Arg	Ser	Ser	Phe	Thr	Val	Gln	
			250				255				260				265	
GAC	CTT	AAA	CCT	TTT	ACA	GAA	TAT	GTG	TTT	AGG	ATT	CGC	TGT	ATG	AAG	1152
Asp	Leu	Lys	Pro	Phe	Thr	Glu	Tyr	Val	Phe	Arg	Ile	Arg	Cys	Met	Lys	
				270					275					280		
GAA	GAT	GGT	AAG	GGA	TAC	TGG	AGT	GAC	TGG	AGT	GAA	GAA	GCA	AGT	GGG	1200
Glu	Asp	Gly	Lys	Gly	Tyr	Trp	Ser	Asp	Trp	Ser	Glu	Glu	Ala	Ser	Gly	
			285					290					295			
ATC	ACC	TAT	GAA	GAT	AGA	CCA	TCT	AAA	GCA	CCA	AGT	TTC	TGG	TAT	AAA	1248
Ile	Thr	Tyr	Glu	Asp	Arg	Pro	Ser	Lys	Ala	Pro	Ser	Phe	Trp	Tyr	Lys	
			300				305					310				
ATA	GAT	CCA	TCC	CAT	ACT	CAA	GGC	TAC	AGA	ACT	GTA	CAA	CTC	GTG	TGG	1296
Ile	Asp	Pro	Ser	His	Thr	Gln	Gly	Tyr	Arg	Thr	Val	Gln	Leu	Val	Trp	
			315				320				325					
AAG	ACA	TTG	CCT	CCT	TTT	GAA	GCC	AAT	GGA	AAA	ATC	TTG	GAT	TAT	GAA	1344
Lys	Thr	Leu	Pro	Pro	Phe	Glu	Ala	Asn	Gly	Lys	Ile	Leu	Asp	Tyr	Glu	
			330				335				340				345	
GTG	ACT	CTC	ACA	AGA	TGG	AAA	TCA	CAT	TTA	CAA	AAT	TAC	ACA	GTT	AAT	1392
Val	Thr	Leu	Thr	Arg	Trp	Lys	Ser	His	Leu	Gln	Asn	Tyr	Thr	Val	Asn	
				350					355					360		

50

GCC	ACA	AAA	CTG	ACA	GTA	AAT	CTC	ACA	AAT	GAT	CGC	TAT	CTA	GCA	ACC	1440
Ala	Thr	Lys	Leu	Thr	Val	Asn	Leu	Thr	Asn	Asp	Arg	Tyr	Leu	Ala	Thr	
			365					370					375			
CTA	ACA	GTA	AGA	AAT	CTT	GTT	GGC	AAA	TCA	GAT	GCA	GCT	GTT	TTA	ACT	1488
Leu	Thr	Val	Arg	Asn	Leu	Val	Gly	Lys	Ser	Asp	Ala	Ala	Val	Leu	Thr	
			380				385					390				
ATC	CCT	GCC	TGT	GAC	TTT	CAA	GCT	ACT	CAC	CCT	GTA	ATG	GAT	CTT	AAA	1536
Ile	Pro	Ala	Cys	Asp	Phe	Gln	Ala	Thr	His	Pro	Val	Met	Asp	Leu	Lys	
	395					400				405						
GCA	TTC	CCC	AAA	GAT	AAC	ATG	CTT	TGG	GTG	GAA	TGG	ACT	ACT	CCA	AGG	1584
Ala	Phe	Pro	Lys	Asp	Asn	Met	Leu	Trp	Val	Glu	Trp	Thr	Thr	Pro	Arg	
410					415					420					425	
GAA	TCT	GTA	AAG	AAA	TAT	ATA	CTT	GAG	TGG	TGT	GTG	TTA	TCA	GAT	AAA	1632
Glu	Ser	Val	Lys	Lys	Tyr	Ile	Leu	Glu	Trp	Cys	Val	Leu	Ser	Asp	Lys	
			430					435						440		
GCA	CCC	TGT	ATC	ACA	GAC	TGG	CAA	CAA	GAA	GAT	GGT	ACC	GTG	CAT	CGC	1680
Ala	Pro	Cys	Ile	Thr	Asp	Trp	Gln	Gln	Glu	Asp	Gly	Thr	Val	His	Arg	
			445				450						455			
ACC	TAT	TTA	AGA	GGG	AAC	TTA	GCA	GAG	AGC	AAA	TGC	TAT	TTG	ATA	ACA	1728
Thr	Tyr	Leu	Arg	Gly	Asn	Leu	Ala	Glu	Ser	Lys	Cys	Tyr	Leu	Ile	Thr	
		460				465						470				
GTT	ACT	CCA	GTA	TAT	GCT	GAT	GGA	CCA	GGA	AGC	CCT	GAA	TCC	ATA	AAG	1776
Val	Thr	Pro	Val	Tyr	Ala	Asp	Gly	Pro	Gly	Ser	Pro	Glu	Ser	Ile	Lys	
	475				480					485						
GCA	TAC	CTT	AAA	CAA	GCT	CCA	CCT	TCC	AAA	GGA	CCT	ACT	GTT	CGG	ACA	1824
Ala	Tyr	Leu	Lys	Gln	Ala	Pro	Pro	Ser	Lys	Gly	Pro	Thr	Val	Arg	Thr	
490				495						500					505	
AAA	AAA	GTA	GGG	AAA	AAC	GAA	GCT	GTC	TTA	GAG	TGG	GAC	CAA	CTT	CCT	1872
Lys	Lys	Val	Gly	Lys	Asn	Glu	Ala	Val	Leu	Glu	Trp	Asp	Gln	Leu	Pro	
			510					515						520		
GTT	GAT	GTT	CAG	AAT	GGA	TTT	ATC	AGA	AAT	TAT	ACT	ATA	TTT	TAT	AGA	1920
Val	Asp	Val	Gln	Asn	Gly	Phe	Ile	Arg	Asn	Tyr	Thr	Ile	Phe	Tyr	Arg	
			525					530					535			
ACC	ATC	ATT	GGA	AAT	GAA	ACT	GCT	GTG	AAT	GTG	GAT	TCT	TCC	CAC	ACA	1968
Thr	Ile	Ile	Gly	Asn	Glu	Thr	Ala	Val	Asn	Val	Asp	Ser	Ser	His	Thr	
		540				545						550				
GAA	TAT	ACA	TTG	TCC	TCT	TTG	ACT	AGT	GAC	ACA	TTG	TAC	ATG	GTA	CGA	2016
Glu	Tyr	Thr	Leu	Ser	Ser	Leu	Thr	Ser	Asp	Thr	Leu	Tyr	Met	Val	Arg	
	555					560					565					
ATG	GCA	GCA	TAC	ACA	GAT	GAA	GGT	GGG	AAG	GAT	GGT	CCA	GAA	TTC	ACT	2064
Met	Ala	Ala	Tyr	Thr	Asp	Glu	Gly	Gly	Lys	Asp	Gly	Pro	Glu	Phe	Thr	
570					575					580					585	
TTT	ACT	ACC	CCA	AAG	TTT	GCT	CAA	GGA	GAA	ATT	GAA	GCC	ATA	GTC	GTG	2112
Phe	Thr	Thr	Pro	Lys	Phe	Ala	Gln	Gly	Glu	Ile	Glu	Ala	Ile	Val	Val	
			590					595						600		

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CCT GTT TGC TTA GCA TTC CTA TTG ACA ACT CTT CTG GGA GTG CTG TTC	2160
Pro Val Cys Leu Ala Phe Leu Leu Thr Thr Leu Leu Gly Val Leu Phe	
605 610 615	
TGC TTT AAT AAG CGA GAC CTA ATT AAA AAA CAC ATC TGG CCT AAT GTT	2208
Cys Phe Asn Lys Arg Asp Leu Ile Lys Lys His Ile Trp Pro Asn Val	
620 625 630	
CCA GAT CCT TCA AAG AGT CAT ATT GCC CAG TGG TCA CCT CAC ACT CCT	2256
Pro Asp Pro Ser Lys Ser His Ile Ala Gln Trp Ser Pro His Thr Pro	
635 640 645	
CCA AGG CAC AAT TTT AAT TCA AAA GAT CAA ATG TAT TCA GAT GGC AAT	2304
Pro Arg His Asn Phe Asn Ser Lys Asp Gln Met Tyr Ser Asp Gly Asn	
650 655 660 665	
TTC ACT GAT GTA AGT GTT GTG GAA ATA GAA GCA AAT GAC AAA AAG CCT	2352
Phe Thr Asp Val Ser Val Val Glu Ile Glu Ala Asn Asp Lys Lys Pro	
670 675 680	
TTT CCA GAA GAT CTG AA	2369
Phe Pro Glu Asp Leu	
685	

## (2) INFORMATION FOR SEQ ID NO:2:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 708 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Leu Thr Leu Gln Thr Trp Leu Val Gln Ala Leu Phe Ile Phe Leu	
-22 -20 -15 -10	
Thr Thr Glu Ser Thr Gly Glu Leu Leu Asp Pro Cys Gly Tyr Ile Ser	
-5 1 5 10	
Pro Glu Ser Pro Val Val Gln Leu His Ser Asn Phe Thr Ala Val Cys	
15 20 25	
Val Leu Lys Glu Lys Cys Met Asp Tyr Phe His Val Asn Ala Asn Tyr	
30 35 40	
Ile Val Trp Lys Thr Asn His Phe Thr Ile Pro Lys Glu Gln Tyr Thr	
45 50 55	
Ile Ile Asn Arg Thr Ala Ser Ser Val Thr Phe Thr Asp Ile Ala Ser	
60 65 70	
Leu Asn Ile Gln Leu Thr Cys Asn Ile Leu Thr Phe Gly Gln Leu Glu	
75 80 85 90	
Gln Asn Val Tyr Gly Ile Thr Ile Ile Ser Gly Leu Pro Pro Glu Lys	
95 100 105	

52

Pro Lys Asn Leu Ser Cys Ile Val Asn Glu Gly Lys Lys Met Arg Cys  
 110 115 120

Glu Trp Asp Gly Gly Arg Glu Thr His Leu Glu Thr Asn Phe Thr Leu  
 125 130 135

Lys Ser Glu Trp Ala Thr His Lys Phe Ala Asp Cys Lys Ala Lys Arg  
 140 145 150

Asp Thr Pro Thr Ser Cys Thr Val Asp Tyr Ser Thr Val Tyr Phe Val  
 155 160 165 170

Asn Ile Glu Val Trp Val Glu Ala Glu Asn Ala Leu Gly Lys Val Thr  
 175 180 185

Ser Asp His Ile Asn Phe Asp Pro Val Tyr Lys Val Lys Pro Asn Pro  
 190 195 200

Pro His Asn Leu Ser Val Ile Asn Ser Glu Glu Leu Ser Ser Ile Leu  
 205 210 215

Lys Leu Thr Trp Thr Asn Pro Ser Ile Lys Ser Val Ile Ile Leu Lys  
 220 225 230

Tyr Asn Ile Gln Tyr Arg Thr Lys Asp Ala Ser Thr Trp Ser Gln Ile  
 235 240 245 250

Pro Pro Glu Asp Thr Ala Ser Thr Arg Ser Ser Phe Thr Val Gln Asp  
 255 260 265

Leu Lys Pro Phe Thr Glu Tyr Val Phe Arg Ile Arg Cys Met Lys Glu  
 270 275 280

Asp Gly Lys Gly Tyr Trp Ser Asp Trp Ser Glu Glu Ala Ser Gly Ile  
 285 290 295

Thr Tyr Glu Asp Arg Pro Ser Lys Ala Pro Ser Phe Trp Tyr Lys Ile  
 300 305 310

Asp Pro Ser His Thr Gln Gly Tyr Arg Thr Val Gln Leu Val Trp Lys  
 315 320 325 330

Thr Leu Pro Pro Phe Glu Ala Asn Gly Lys Ile Leu Asp Tyr Glu Val  
 335 340 345

Thr Leu Thr Arg Trp Lys Ser His Leu Gln Asn Tyr Thr Val Asn Ala  
 350 355 360

Thr Lys Leu Thr Val Asn Leu Thr Asn Asp Arg Tyr Leu Ala Thr Leu  
 365 370 375

Thr Val Arg Asn Leu Val Gly Lys Ser Asp Ala Ala Val Leu Thr Ile  
 380 385 390

Pro Ala Cys Asp Phe Gln Ala Thr His Pro Val Met Asp Leu Lys Ala  
 395 400 405 410

Phe Pro Lys Asp Asn Met Leu Trp Val Glu Trp Thr Thr Pro Arg Glu  
 415 420 425

53

Ser Val Lys Lys Tyr Ile Leu Glu Trp Cys Val Leu Ser Asp Lys Ala  
 430 435 440  
 Pro Cys Ile Thr Asp Trp Gln Gln Glu Asp Gly Thr Val His Arg Thr  
 445 450 455  
 Tyr Leu Arg Gly Asn Leu Ala Glu Ser Lys Cys Tyr Leu Ile Thr Val  
 460 465 470  
 Thr Pro Val Tyr Ala Asp Gly Pro Gly Ser Pro Glu Ser Ile Lys Ala  
 475 480 485 490  
 Tyr Leu Lys Gln Ala Pro Pro Ser Lys Gly Pro Thr Val Arg Thr Lys  
 495 500 505  
 Lys Val Gly Lys Asn Glu Ala Val Leu Glu Trp Asp Gln Leu Pro Val  
 510 515 520  
 Asp Val Gln Asn Gly Phe Ile Arg Asn Tyr Thr Ile Phe Tyr Arg Thr  
 525 530 535  
 Ile Ile Gly Asn Glu Thr Ala Val Asn Val Asp Ser Ser His Thr Glu  
 540 545 550  
 Tyr Thr Leu Ser Ser Leu Thr Ser Asp Thr Leu Tyr Met Val Arg Met  
 555 560 565 570  
 Ala Ala Tyr Thr Asp Glu Gly Gly Lys Asp Gly Pro Glu Phe Thr Phe  
 575 580 585  
 Thr Thr Pro Lys Phe Ala Gln Gly Glu Ile Glu Ala Ile Val Val Pro  
 590 595 600  
 Val Cys Leu Ala Phe Leu Leu Thr Thr Leu Leu Gly Val Leu Phe Cys  
 605 610 615  
 Phe Asn Lys Arg Asp Leu Ile Lys Lys His Ile Trp Pro Asn Val Pro  
 620 625 630  
 Asp Pro Ser Lys Ser His Ile Ala Gln Trp Ser Pro His Thr Pro Pro  
 635 640 645 650  
 Arg His Asn Phe Asn Ser Lys Asp Gln Met Tyr Ser Asp Gly Asn Phe  
 655 660 665  
 Thr Asp Val Ser Val Val Glu Ile Glu Ala Asn Asp Lys Lys Pro Phe  
 670 675 680  
 Pro Glu Asp Leu  
 685

## (2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 705 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

54

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vii) IMMEDIATE SOURCE:

(B) CLONE: hIgG1Fc

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..699

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GAG CCC AGA TCT TGT GAC AAA ACT CAC ACA TGC CCA CCG TGC CCA GCA	48
Glu Pro Arg Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala	
1 5 10 15	
CCT GAA CTC CTG GGG GGA CCG TCA GTC TTC CTC TTC CCC CCA AAA CCC	96
Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro	
20 25 30	
AAG GAC ACC CTC ATG ATC TCC CGG ACC CCT GAG GTC ACA TGC GTG GTG	144
Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val	
35 40 45	
GTG GAC GTG AGC CAC GAA GAC CCT GAG GTC AAG TTC AAC TGG TAC GTG	192
Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val	
50 55 60	
GAC GGC GTG GAG GTG CAT AAT GCC AAG ACA AAG CCG CGG GAG GAG CAG	240
Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln	
65 70 75 80	
TAC AAC AGC ACG TAC CGG GTG GTC AGC GTC CTC ACC GTC CTG CAC CAG	288
Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln	
85 90 95	
GAC TGG CTG AAT GGC AAG GAC TAC AAG TGC AAG GTC TCC AAC AAA GCC	336
Asp Trp Leu Asn Gly Lys Asp Tyr Lys Cys Lys Val Ser Asn Lys Ala	
100 105 110	
CTC CCA GCC CCC ATG CAG AAA ACC ATC TCC AAA GCC AAA GGG CAG CCC	384
Leu Pro Ala Pro Met Gln Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro	
115 120 125	
CGA GAA CCA CAG GTG TAC ACC CTG CCC CCA TCC CGG GAT GAG CTG ACC	432
Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr	
130 135 140	
AAG AAC CAG GTC AGC CTG ACC TGC CTG GTC AAA GGC TTC TAT CCC AGG	480
Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Arg	
145 150 155 160	
CAC ATC GCC GTG GAG TGG GAG AGC AAT GGG CAG CCG GAG AAC AAC TAC	528
His Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr	
165 170 175	

55

AAG ACC ACG CCT CCC GTG CTG GAC TCC GAC GGC TCC TTC TTC CTC TAC	576
Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr	
180 185 190	
AGC AAG CTC ACC GTG GAC AAG AGC AGG TGG CAG CAG GGG AAC GTC TTC	624
Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe	
195 200 205	
TCA TGC TCC GTG ATG CAT GAG GCT CTG CAC AAC CAC TAC ACG CAG AAG	672
Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys	
210 215 220	
AGC CTC TCC CTG TCT CCG GGT AAA TGAAGTAGT	705
Ser Leu Ser Leu Ser Pro Gly Lys	
225 230	

## (2) INFORMATION FOR SEQ ID NO:4:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 232 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Glu Pro Arg Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala	
1 5 10 15	
Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro	
20 25 30	
Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val	
35 40 45	
Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val	
50 55 60	
Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln	
65 70 75 80	
Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln	
85 90 95	
Asp Trp Leu Asn Gly Lys Asp Tyr Lys Cys Lys Val Ser Asn Lys Ala	
100 105 110	
Leu Pro Ala Pro Met Gln Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro	
115 120 125	
Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr	
130 135 140	
Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Arg	
145 150 155 160	
His Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr	
165 170 175	

56

Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr  
                   180                  185                  190  
 Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe  
                   195                  200                  205  
 Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys  
                   210                  215                  220  
 Ser Leu Ser Leu Ser Pro Gly Lys  
                   225                  230

## (2) INFORMATION FOR SEQ ID NO:5:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3182 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: cDNA to mRNA

## (iii) HYPOTHETICAL: NO

## (iv) ANTI-SENSE: NO

## (v) FRAGMENT TYPE: N-terminal

## (vi) ORIGINAL SOURCE:

- (F) TISSUE TYPE: human placenta

## (vii) IMMEDIATE SOURCE:

- (B) CLONE: PHLIFR-65

## (ix) FEATURE:

- (A) NAME/KEY: mat\_peptide
- (B) LOCATION: 311..3182

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 179..3182

## (ix) FEATURE:

- (A) NAME/KEY: sig\_peptide
- (B) LOCATION: 179..310

## (ix) FEATURE:

- (A) NAME/KEY: mat\_peptide
- (B) LOCATION: 311..3182

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

AGATCTTGGA ACGAGACGAC CTGCTCTCTC TCCCAGAACG TGTCTCTGCT GCAAGGCACC 60  
 GGGCCCTTTC GCTCTGCAGA ACTGCACTTG CAAGACCATT ATCAACTCCT AATCCCA3CT 120  
 CAGAAAGGGA GCCTCTGCGA CTCATTCATC GCCCTCCAGG ACTGACTGCA TTGCACAG 178

57

ATG	ATG	GAT	ATT	TAC	GTA	TGT	TTG	AAA	CGA	CCA	TCC	TGG	ATG	GTG	GAC	226
Met	Met	Asp	Ile	Tyr	Val	Cys	Leu	Lys	Arg	Pro	Ser	Trp	Met	Val	Asp	
-44				-40					-35					-30		
AAT	AAA	AGA	ATG	AGG	ACT	GCT	TCA	AAT	TTC	CAG	TGG	CTG	TTA	TCA	ACA	274
Asn	Lys	Arg	Met	Arg	Thr	Ala	Ser	Asn	Phe	Gln	Trp	Leu	Leu	Ser	Thr	
			-25					-20					-15			
TTT	ATT	CTT	CTA	TAT	CTA	ATG	AAT	CAA	GTA	AAT	AGC	CAG	AAA	AAG	GGG	322
Phe	Ile	Leu	Leu	Tyr	Leu	Met	Asn	Gln	Val	Asn	Ser	Gln	Lys	Lys	Gly	
		-10					-5						1			
GCT	CCT	CAT	GAT	TTG	AAG	TGT	GTA	ACT	AAC	AAT	TTG	CAA	GTG	TGG	AAC	370
Ala	Pro	His	Asp	Leu	Lys	Cys	Val	Thr	Asn	Asn	Leu	Gln	Val	Trp	Asn	
5					10					15					20	
TGT	TCT	TGG	AAA	GCA	CCC	TCT	GGA	ACA	GGC	CGT	GGT	ACT	GAT	TAT	GAA	418
Cys	Ser	Trp	Lys	Ala	Pro	Ser	Gly	Thr	Gly	Arg	Gly	Thr	Asp	Tyr	Glu	
			25						30					35		
GTT	TGC	ATT	GAA	AAC	AGG	TCC	CGT	TCT	TGT	TAT	CAG	TTG	GAG	AAA	ACC	466
Val	Cys	Ile	Glu	Asn	Arg	Ser	Arg	Ser	Cys	Tyr	Gln	Leu	Glu	Lys	Thr	
			40					45					50			
AGT	ATT	AAA	ATT	CCA	GCT	CTT	TCA	CAT	GGT	GAT	TAT	GAA	ATA	ACA	ATA	514
Ser	Ile	Lys	Ile	Pro	Ala	Leu	Ser	His	Gly	Asp	Tyr	Glu	Ile	Thr	Ile	
		55					60					65				
AAT	TCT	CTA	CAT	GAT	TTT	GGA	AGT	TCT	ACA	AGT	AAA	TTC	ACA	CTA	AAT	562
Asn	Ser	Leu	His	Asp	Phe	Gly	Ser	Ser	Thr	Ser	Lys	Phe	Thr	Leu	Asn	
		70				75					80					
GAA	CAA	AAC	GTT	TCC	TTA	ATT	CCA	GAT	ACT	CCA	GAG	ATC	TTG	AAT	TTG	610
Glu	Gln	Asn	Val	Ser	Leu	Ile	Pro	Asp	Thr	Pro	Glu	Ile	Leu	Asn	Leu	
85					90					95					100	
TCT	GCT	GAT	TTC	TCA	ACC	TCT	ACA	TTA	TAC	CTA	AAG	TGG	AAC	GAC	AGG	658
Ser	Ala	Asp	Phe	Ser	Thr	Ser	Thr	Leu	Tyr	Leu	Lys	Trp	Asn	Asp	Arg	
				105					110					115		
GGT	TCA	GTT	TTT	CCA	CAC	CGC	TCA	AAT	GTT	ATC	TGG	GAA	ATT	AAA	GTT	706
Gly	Ser	Val	Phe	Pro	His	Arg	Ser	Asn	Val	Ile	Trp	Glu	Ile	Lys	Val	
			120					125					130			
CTA	CGT	AAA	GAG	AGT	ATG	GAG	CTC	GTA	AAA	TTA	GTG	ACC	CAC	AAC	ACA	754
Leu	Arg	Lys	Glu	Ser	Met	Glu	Leu	Val	Lys	Leu	Val	Thr	His	Asn	Thr	
		135					140					145				
ACT	CTG	AAT	GGC	AAA	GAT	ACA	CTT	CAT	CAC	TGG	AGT	TGG	GCC	TCA	GAT	802
Thr	Leu	Asn	Gly	Lys	Asp	Thr	Leu	His	His	Trp	Ser	Trp	Ala	Ser	Asp	
		150				155					160					
ATG	CCC	TTG	GAA	TGT	GCC	ATT	CAT	TTT	GTG	GAA	ATT	AGA	TGC	TAC	ATT	850
Met	Pro	Leu	Glu	Cys	Ala	Ile	His	Phe	Val	Glu	Ile	Arg	Cys	Tyr	Ile	
165					170					175					180	
GAC	AAT	CTT	CAT	TTT	TCT	GGT	CTC	GAA	GAG	TGG	AGT	GAC	TGG	AGC	CCT	898
Asp	Asn	Leu	His	Phe	Ser	Gly	Leu	Glu	Glu	Trp	Ser	Asp	Trp	Ser	Pro	
				185					190					195		

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GTG	AAG	AAC	ATT	TCT	TGG	ATA	CCT	GAT	TCT	CAG	ACT	AAG	GTT	TTT	CCT	946
Val	Lys	Asn	Ile	Ser	Trp	Ile	Pro	Asp	Ser	Gln	Thr	Lys	Val	Phe	Pro	
			200					205					210			
CAA	GAT	AAA	GTG	ATA	CTT	GTA	GGC	TCA	GAC	ATA	ACA	TTT	TGT	TGT	GTG	994
Gln	Asp	Lys	Val	Ile	Leu	Val	Gly	Ser	Asp	Ile	Thr	Phe	Cys	Cys	Val	
		215					220					225				
AGT	CAA	GAA	AAA	GTG	TTA	TCA	GCA	CTG	ATT	GGC	CAT	ACA	AAC	TGC	CCC	1042
Ser	Gln	Glu	Lys	Val	Leu	Ser	Ala	Leu	Ile	Gly	His	Thr	Asn	Cys	Pro	
	230					235					240					
TTG	ATC	CAT	CTT	GAT	GGG	GAA	AAT	GTT	GCA	ATC	AAG	ATT	CGT	AAT	ATT	1090
Leu	Ile	His	Leu	Asp	Gly	Glu	Asn	Val	Ala	Ile	Lys	Ile	Arg	Asn	Ile	
245					250					255					260	
TCT	GTT	TCT	GCA	AGT	AGT	GGA	ACA	AAT	GTA	GTT	TTT	ACA	ACC	GAA	GAT	1138
Ser	Val	Ser	Ala	Ser	Ser	Gly	Thr	Asn	Val	Val	Phe	Thr	Thr	Glu	Asp	
				265					270					275		
AAC	ATA	TTT	GGA	ACC	GTT	ATT	TTT	GCT	GGA	TAT	CCA	CCA	GAT	ACT	CCT	1186
Asn	Ile	Phe	Gly	Thr	Val	Ile	Phe	Ala	Gly	Tyr	Pro	Pro	Asp	Thr	Pro	
			280					285					290			
CAA	CAA	CTG	AAT	TGT	GAG	ACA	CAT	GAT	TTA	AAA	GAA	ATT	ATA	TGT	AGT	1234
Gln	Gln	Leu	Asn	Cys	Glu	Thr	His	Asp	Leu	Lys	Glu	Ile	Ile	Cys	Ser	
		295					300					305				
TGG	AAT	CCA	GGA	AGG	GTG	ACA	GCG	TTG	GTG	GGC	CCA	CGT	GCT	ACA	AGC	1282
Trp	Asn	Pro	Gly	Arg	Val	Thr	Ala	Leu	Val	Gly	Pro	Arg	Ala	Thr	Ser	
	310					315					320					
TAC	ACT	TTA	GTT	GAA	AGT	TTT	TCA	GGA	AAA	TAT	GTT	AGA	CTT	AAA	AGA	1330
Tyr	Thr	Leu	Val	Glu	Ser	Phe	Ser	Gly	Lys	Tyr	Val	Arg	Leu	Lys	Arg	
325					330					335					340	
GCT	GAA	GCA	CCT	ACA	AAC	GAA	AGC	TAT	CAA	TTA	TTA	TTT	CAA	ATG	CTT	1378
Ala	Glu	Ala	Pro	Thr	Asn	Glu	Ser	Tyr	Gln	Leu	Leu	Phe	Gln	Met	Leu	
				345					350					355		
CCA	AAT	CAA	GAA	ATA	TAT	AAT	TTT	ACT	TTG	AAT	GCT	CAC	AAT	CCG	CTG	1426
Pro	Asn	Gln	Glu	Ile	Tyr	Asn	Phe	Thr	Leu	Asn	Ala	His	Asn	Pro	Leu	
			360					365					370			
GGT	CGA	TCA	CAA	TCA	ACA	ATT	TTA	GTT	AAT	ATA	ACT	GAA	AAA	GTT	TAT	1474
Gly	Arg	Ser	Gln	Ser	Thr	Ile	Leu	Val	Asn	Ile	Thr	Glu	Lys	Val	Tyr	
		375					380					385				
CCC	CAT	ACT	CCT	ACT	TCA	TTC	AAA	GTG	AAG	GAT	ATT	AAT	TCA	ACA	GCT	1522
Pro	His	Thr	Pro	Thr	Ser	Phe	Lys	Val	Lys	Asp	Ile	Asn	Ser	Thr	Ala	
	390					395					400					
GTT	AAA	CTT	TCT	TGG	CAT	TTA	CCA	GGC	AAC	TTT	GCA	AAG	ATT	AAT	TTT	1570
Val	Lys	Leu	Ser	Trp	His	Leu	Pro	Gly	Asn	Phe	Ala	Lys	Ile	Asn	Phe	
405					410					415					420	
TTA	TGT	GAA	ATT	GAA	ATT	AAG	AAA	TCT	AAT	TCA	GTA	CAA	GAG	CAG	CGG	1616
Leu	Cys	Glu	Ile	Glu	Ile	Lys	Lys	Ser	Asn	Ser	Val	Gln	Glu	Gln	Arg	
				425					430					435		

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AAT GTC ACA ATC AAA GGA GTA GAA AAT TCA AGT TAT CTT GTT GCT CTG	1666
Asn Val Thr Ile Lys Gly Val Glu Asn Ser Ser Tyr Leu Val Ala Leu	
440 445 450	
GAC AAG TTA AAT CCA TAC ACT CTA TAT ACT TTT CGG ATT CGT TGT TCT	1714
Asp Lys Leu Asn Pro Tyr Thr Leu Tyr Thr Phe Arg Ile Arg Cys Ser	
455 460 465	
ACT GAA ACT TTC TGG AAA TGG AGC AAA TGG AGC AAT AAA AAA CAA CAT	1762
Thr Glu Thr Phe Trp Lys Trp Ser Lys Trp Ser Asn Lys Lys Gln His	
470 475 480	
TTA ACA ACA GAA GCC AGT CCT TCA AAG GGG CCT GAT ACT TGG AGA GAG	1810
Leu Thr Thr Glu Ala Ser Pro Ser Lys Gly Pro Asp Thr Trp Arg Glu	
485 490 495 500	
TGG AGT TCT GAT GGA AAA AAT TTA ATA ATC TAT TGG AAG CCT TTA CCC	1858
Trp Ser Ser Asp Gly Lys Asn Leu Ile Ile Tyr Trp Lys Pro Leu Pro	
505 510 515	
ATT AAT GAA GCT AAT GGA AAA ATA CTT TCC TAC AAT GTA TCG TGT TCA	1906
Ile Asn Glu Ala Asn Gly Lys Ile Leu Ser Tyr Asn Val Ser Cys Ser	
520 525 530	
TCA GAT GAG GAA ACA CAG TCC CTT TCT GAA ATC CCT GAT CCT CAG CAC	1954
Ser Asp Glu Glu Thr Gln Ser Leu Ser Glu Ile Pro Asp Pro Gln His	
535 540 545	
AAA GCA GAG ATA CGA CTT GAT AAG AAT GAC TAC ATC ATC AGC GTA GTG	2002
Lys Ala Glu Ile Arg Leu Asp Lys Asn Asp Tyr Ile Ile Ser Val Val	
550 555 560	
GCT AAA AAT TCT GTG GGC TCA TCA CCA CCT TCC AAA ATA GCG AGT ATG	2050
Ala Lys Asn Ser Val Gly Ser Ser Pro Pro Ser Lys Ile Ala Ser Met	
565 570 575 580	
GAA ATT CCA AAT GAT GAT CTC AAA ATA GAA CAA GTT GTT GGG ATG GGA	2098
Glu Ile Pro Asn Asp Asp Leu Lys Ile Glu Gln Val Val Gly Met Gly	
585 590 595	
AAG GGG ATT CTC CTC ACC TGG CAT TAC GAC CCC AAC ATG ACT TGC GAC	2146
Lys Gly Ile Leu Leu Thr Trp His Tyr Asp Pro Asn Met Thr Cys Asp	
600 605 610	
TAC GTC ATT AAG TGG TGT AAC TCG TCT CGG TCG GAA CCA TGC CTT ATG	2194
Tyr Val Ile Lys Trp Cys Asn Ser Ser Arg Ser Glu Pro Cys Leu Met	
615 620 625	
GAC TGG AGA AAA GTT CCC TCA AAC AGC ACT GAA ACT GTA ATA GAA TCT	2242
Asp Trp Arg Lys Val Prp Ser Asn Ser Thr Glu Thr Val Ile Glu Ser	
630 635 640	
GAT GAG TTT CGA CCA GGT ATA AGA TAT AAT TTT TTC CTG TAT GGA TGC	2290
Asp Glu Phe Arg Pro Gly Ile Arg Tyr Asn Phe Phe Leu Tyr Gly Cys	
645 650 655 660	
AGA AAT CAA GGA TAT CAA TTA TTA CGC TCC ATG ATT GGA TAT ATA GAA	2338
Arg Asn Gln Gly Tyr Gln Leu Leu Arg Ser Met Ile Gly Tyr Ile Glu	
665 670 675	

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GAA	TTG	GCT	CCC	ATT	GTT	GCA	CCA	AAT	TTT	ACT	GTT	GAG	GAT	ACT	TCT	2386
Glu	Leu	Ala	Pro	Ile	Val	Ala	Pro	Asn	Phe	Thr	Val	Glu	Asp	Thr	Ser	
			680					685					690			
GCA	GAT	TCG	ATA	TTA	GTA	AAA	TGG	GAA	GAC	ATT	CCT	GTG	GAA	GAA	CTT	2434
Ala	Asp	Ser	Ile	Leu	Val	Lys	Trp	Glu	Asp	Ile	Pro	Val	Glu	Glu	Leu	
		695					700					705				
AGA	GGC	TTT	TTA	AGA	GGA	TAT	TTG	TTT	TAC	TTT	GGA	AAA	GGA	GAA	AGA	2482
Arg	Gly	Phe	Leu	Arg	Gly	Tyr	Leu	Phe	Tyr	Phe	Gly	Lys	Gly	Glu	Arg	
	710					715					720					
GAC	ACA	TCT	AAG	ATG	AGG	GTT	TTA	GAA	TCA	GGT	CGT	TCT	GAC	ATA	AAA	2530
Asp	Thr	Ser	Lys	Met	Arg	Val	Leu	Glu	Ser	Gly	Arg	Ser	Asp	Ile	Lys	
	725				730					735					740	
GTT	AAG	AAT	ATT	ACT	GAC	ATA	TCC	CAG	AAG	ACA	CTG	AGA	ATT	GCT	GAT	2578
Val	Lys	Asn	Ile	Thr	Asp	Ile	Ser	Gln	Lys	Thr	Leu	Arg	Ile	Ala	Asp	
			745					750						755		
CTT	CAA	GGT	AAA	ACA	AGT	TAC	CAC	CTG	GTC	TTG	CGA	GCC	TAT	ACA	GAT	2626
Leu	Gln	Gly	Lys	Thr	Ser	Tyr	His	Leu	Val	Leu	Arg	Ala	Tyr	Thr	Asp	
		760					765						770			
GGT	GGA	GTG	GGC	CCG	GAG	AAG	AGT	ATG	TAT	GTG	GTG	ACA	AAG	GAA	AAT	2674
Gly	Gly	Val	Gly	Pro	Glu	Lys	Ser	Met	Tyr	Val	Val	Thr	Lys	Glu	Asn	
		775				780						785				
TCT	GTG	GGA	TTA	ATT	ATT	GCC	ATT	CTC	ATC	CCA	GTG	GCA	GTG	GCT	GTC	2722
Ser	Val	Gly	Leu	Ile	Ile	Ala	Ile	Leu	Ile	Pro	Val	Ala	Val	Ala	Val	
	790					795					800					
ATT	GTT	GGA	GTG	GTG	ACA	AGT	ATC	CTT	TGC	TAT	CGG	AAA	CGA	GAA	TGG	2770
Ile	Val	Gly	Val	Val	Thr	Ser	Ile	Leu	Cys	Tyr	Arg	Lys	Arg	Glu	Trp	
	805				810					815				820		
ATT	AAA	GAA	ACC	TTC	TAC	CCT	GAT	ATT	CCA	AAT	CCA	GAA	AAC	TGT	AAA	2818
Ile	Lys	Glu	Thr	Phe	Tyr	Pro	Asp	Ile	Pro	Asn	Pro	Glu	Asn	Cys	Lys	
			825					830						835		
GCA	TTA	CAG	TTT	CAA	AAG	AGT	GTC	TGT	GAG	GGA	AGC	AGT	GCT	CTT	AAA	2866
Ala	Leu	Gln	Phe	Gln	Lys	Ser	Val	Cys	Glu	Gly	Ser	Ser	Ala	Leu	Lys	
		840					845						850			
ACA	TTG	GAA	ATG	AAT	CCT	TGT	ACC	CCA	AAT	AAT	GTT	GAG	GTT	CTG	GAA	2914
Thr	Leu	Glu	Met	Asn	Pro	Cys	Thr	Pro	Asn	Asn	Val	Glu	Val	Leu	Glu	
		855				860					865					
ACT	CGA	TCA	GCA	TTT	CCT	AAA	ATA	GAA	GAT	ACA	GAA	ATA	ATT	TCC	CCA	2962
Thr	Arg	Ser	Ala	Phe	Pro	Lys	Ile	Glu	Asp	Thr	Glu	Ile	Ile	Ser	Pro	
	870					875					880					
GTA	GCT	GAG	CGT	CCT	GAA	GAT	CGC	TCT	GAT	GCA	GAG	CCT	GAA	AAC	CAT	3010
Val	Ala	Glu	Arg	Pro	Glu	Asp	Arg	Ser	Asp	Glu	Glu	Pro	Glu	Asn	His	
	885				890					895				900		
GTG	GTT	GTG	TCC	TAT	TGT	CCA	CCC	ATC	ATT	GAG	GAA	GAA	ATA	CCA	AAC	3058
Val	Val	Val	Ser	Tyr	Cys	Pro	Pro	Ile	Ile	Glu	Glu	Glu	Ile	Pro	Asn	
			905					910						915		

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CCA	GCC	GCA	GAT	GAA	GCT	GGA	GGG	ACT	GCA	CAG	GTT	ATT	TAC	ATT	GAT	3106
Pro	Ala	Ala	Asp	Glu	Ala	Gly	Gly	Thr	Ala	Gln	Val	Ile	Tyr	Ile	Asp	
			920					925					930			
GTT	CAG	TCG	ATG	TAT	CAG	CCT	CAA	GCA	AAA	CCA	GAA	GAA	AAA	AAA	AAA	3154
Val	Gln	Ser	Met	Tyr	Gln	Pro	Gln	Ala	Lys	Pro	Glu	Glu	Lys	Lys	Lys	
		935					940					945				
AAA	AGC	AGG	TCG	TCT	CGT	TCC	AAG	ATC	T							3182
Lys	Ser	Arg	Ser	Ser	Arg	Ser	Lys	Ile								
		950					955									

## (2) INFORMATION FOR SEQ ID NO:6:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1001 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met	Met	Asp	Ile	Tyr	Val	Cys	Leu	Lys	Arg	Pro	Ser	Trp	Met	Val	Asp	
-44				-40					-35					-30		
Asn	Lys	Arg	Met	Arg	Thr	Ala	Ser	Asn	Phe	Gln	Trp	Leu	Leu	Ser	Thr	
		-25						-20						-15		
Phe	Ile	Leu	Leu	Tyr	Leu	Met	Asn	Gln	Val	Asn	Ser	Gln	Lys	Lys	Gly	
		-10					-5					1				
Ala	Pro	His	Asp	Leu	Lys	Cys	Val	Thr	Asn	Asn	Leu	Gln	Val	Trp	Asn	
5					10					15					20	
Cys	Ser	Trp	Lys	Ala	Pro	Ser	Gly	Thr	Gly	Arg	Gly	Thr	Asp	Tyr	Glu	
			25						30					35		
Val	Cys	Ile	Glu	Asn	Arg	Ser	Arg	Ser	Cys	Tyr	Gln	Leu	Glu	Lys	Thr	
			40					45					50			
Ser	Ile	Lys	Ile	Pro	Ala	Leu	Ser	His	Gly	Asp	Tyr	Glu	Ile	Thr	Ile	
		55					60					65				
Asn	Ser	Leu	His	Asp	Phe	Gly	Ser	Ser	Thr	Ser	Lys	Phe	Thr	Leu	Asn	
		70				75					80					
Glu	Gln	Asn	Val	Ser	Leu	Ile	Pro	Asp	Thr	Pro	Glu	Ile	Leu	Asn	Leu	
		85			90					95					100	
Ser	Ala	Asp	Phe	Ser	Thr	Ser	Thr	Leu	Tyr	Leu	Lys	Trp	Asn	Asp	Arg	
			105						110					115		
Gly	Ser	Val	Phe	Pro	His	Arg	Ser	Asn	Val	Ile	Trp	Glu	Ile	Lys	Val	
			120					125					130			
Leu	Arg	Lys	Glu	Ser	Met	Glu	Leu	Val	Lys	Leu	Val	Thr	His	Asn	Thr	
		135					140					145				

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Thr Leu Asn Gly Lys Asp Thr Leu His His Trp Ser Trp Ala Ser Asp  
 150 155 160  
 Met Pro Leu Glu Cys Ala Ile His Phe Val Glu Ile Arg Cys Tyr Ile  
 165 170 175 180  
 Asp Asn Leu His Phe Ser Gly Leu Glu Glu Trp Ser Asp Trp Ser Pro  
 185 190 195  
 Val Lys Asn Ile Ser Trp Ile Pro Asp Ser Gln Thr Lys Val Phe Pro  
 200 205 210  
 Gln Asp Lys Val Ile Leu Val Gly Ser Asp Ile Thr Phe Cys Cys Val  
 215 220 225  
 Ser Gln Glu Lys Val Leu Ser Ala Leu Ile Gly His Thr Asn Cys Pro  
 230 235 240  
 Leu Ile His Leu Asp Gly Glu Asn Val Ala Ile Lys Ile Arg Asn Ile  
 245 250 255 260  
 Ser Val Ser Ala Ser Ser Gly Thr Asn Val Val Phe Thr Thr Glu Asp  
 265 270 275  
 Asn Ile Phe Gly Thr Val Ile Phe Ala Gly Tyr Pro Pro Asp Thr Pro  
 280 285 290  
 Gln Gln Leu Asn Cys Glu Thr His Asp Leu Lys Glu Ile Ile Cys Ser  
 295 300 305  
 Trp Asn Pro Gly Arg Val Thr Ala Leu Val Gly Pro Arg Ala Thr Ser  
 310 315 320  
 Tyr Thr Leu Val Glu Ser Phe Ser Gly Lys Tyr Val Arg Leu Lys Arg  
 325 330 335 340  
 Ala Glu Ala Pro Thr Asn Glu Ser Tyr Gln Leu Leu Phe Gln Met Leu  
 345 350 355  
 Pro Asn Gln Glu Ile Tyr Asn Phe Thr Leu Asn Ala His Asn Pro Leu  
 360 365 370  
 Gly Arg Ser Gln Ser Thr Ile Leu Val Asn Ile Thr Glu Lys Val Tyr  
 375 380 385  
 Pro His Thr Pro Thr Ser Phe Lys Val Lys Asp Ile Asn Ser Thr Ala  
 390 395 400  
 Val Lys Leu Ser Trp His Leu Pro Gly Asn Phe Ala Lys Ile Asn Phe  
 405 410 415 420  
 Leu Cys Glu Ile Glu Ile Lys Lys Ser Asn Ser Val Gln Glu Gln Arg  
 425 430 435  
 Asn Val Thr Ile Lys Gly Val Glu Asn Ser Ser Tyr Leu Val Ala Leu  
 440 445 450  
 Asp Lys Leu Asn Pro Tyr Thr Leu Tyr Thr Phe Arg Ile Arg Cys Ser  
 455 460 465

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Thr	Glu	Thr	Phe	Trp	Lys	Trp	Ser	Lys	Trp	Ser	Asn	Lys	Lys	Gln	His
470						475					480				
Leu	Thr	Thr	Glu	Ala	Ser	Pro	Ser	Lys	Gly	Pro	Asp	Thr	Trp	Arg	Glu
485					490					495					500
Trp	Ser	Ser	Asp	Gly	Lys	Asn	Leu	Ile	Ile	Tyr	Trp	Lys	Pro	Leu	Pro
				505					510					515	
Ile	Asn	Glu	Ala	Asn	Gly	Lys	Ile	Leu	Ser	Tyr	Asn	Val	Ser	Cys	Ser
			520					525					530		
Ser	Asp	Glu	Glu	Thr	Gln	Ser	Leu	Ser	Glu	Ile	Pro	Asp	Pro	Gln	His
		535					540					545			
Lys	Ala	Glu	Ile	Arg	Leu	Asp	Lys	Asn	Asp	Tyr	Ile	Ile	Ser	Val	Val
	550					555					560				
Ala	Lys	Asn	Ser	Val	Gly	Ser	Ser	Pro	Pro	Ser	Lys	Ile	Ala	Ser	Met
565					570					575					580
Glu	Ile	Pro	Asn	Asp	Asp	Leu	Lys	Ile	Glu	Gln	Val	Val	Gly	Met	Gly
				585					590					595	
Lys	Gly	Ile	Leu	Leu	Thr	Trp	His	Tyr	Asp	Pro	Asn	Met	Thr	Cys	Asp
			600					605					610		
Tyr	Val	Ile	Lys	Trp	Cys	Asn	Ser	Ser	Arg	Ser	Glu	Pro	Cys	Leu	Met
	615						620					625			
Asp	Trp	Arg	Lys	Val	Pro	Ser	Asn	Ser	Thr	Glu	Thr	Val	Ile	Glu	Ser
	630						635					640			
Asp	Glu	Phe	Arg	Pro	Gly	Ile	Arg	Tyr	Asn	Phe	Phe	Leu	Tyr	Gly	Cys
645					650					655					660
Arg	Asn	Gln	Gly	Tyr	Gln	Leu	Leu	Arg	Ser	Met	Ile	Gly	Tyr	Ile	Glu
				665					670					675	
Glu	Leu	Ala	Pro	Ile	Val	Ala	Pro	Asn	Phe	Thr	Val	Glu	Asp	Thr	Ser
			680					685					690		
Ala	Asp	Ser	Ile	Leu	Val	Lys	Trp	Glu	Asp	Ile	Pro	Val	Glu	Glu	Leu
		695					700					705			
Arg	Gly	Phe	Leu	Arg	Gly	Tyr	Leu	Phe	Tyr	Phe	Gly	Lys	Gly	Glu	Arg
	710					715					720				
Asp	Thr	Ser	Lys	Met	Arg	Val	Leu	Glu	Ser	Gly	Arg	Ser	Asp	Ile	Lys
	725				730					735					740
Val	Lys	Asn	Ile	Thr	Asp	Ile	Ser	Gln	Lys	Thr	Leu	Arg	Ile	Ala	Asp
				745					750					755	
Leu	Gln	Gly	Lys	Thr	Ser	Tyr	His	Leu	Val	Leu	Arg	Ala	Tyr	Thr	Asp
			760					765					770		
Gly	Gly	Val	Gly	Pro	Glu	Lys	Ser	Met	Tyr	Val	Val	Thr	Lys	Glu	Asn
		775					780						785		

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Ser Val Gly Leu Ile Ile Ala Ile Leu Ile Pro Val Ala Val Ala Val  
 790 795 800  
 Ile Val Gly Val Val Thr Ser Ile Leu Cys Tyr Arg Lys Arg Glu Trp  
 805 810 815 820  
 Ile Lys Glu Thr Phe Tyr Pro Asp Ile Pro Asn Pro Glu Asn Cys Lys  
 825 830 835  
 Ala Leu Gln Phe Gln Lys Ser Val Cys Glu Gly Ser Ser Ala Leu Lys  
 840 845 850  
 Thr Leu Glu Met Asn Pro Cys Thr Pro Asn Asn Val Glu Val Leu Glu  
 855 860 865  
 Thr Arg Ser Ala Phe Pro Lys Ile Glu Asp Thr Glu Ile Ile Ser Pro  
 870 875 880  
 Val Ala Glu Arg Pro Glu Asp Arg Ser Asp Ala Glu Pro Glu Asn His  
 885 890 895 900  
 Val Val Val Ser Tyr Cys Pro Pro Ile Ile Glu Glu Glu Ile Pro Asn  
 905 910 915  
 Pro Ala Ala Asp Glu Ala Gly Gly Thr Ala Gln Val Ile Tyr Ile Asp  
 920 925 930  
 Val Gln Ser Met Tyr Gln Pro Gln Ala Lys Pro Glu Glu Lys Lys Lys  
 935 940 945  
 Lys Ser Arg Ser Ser Arg Ser Lys Ile  
 950 955

## (2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 100 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

GATCCGGTGG AGGTGGTTCT GGTGGAGGTG GTTCAGGTGG TGGAGGATCA GGAGGTGGTG 60  
 GATCAGGTGG AGGAGGTTCT GGAGGTGGAG GTTCCGAAT 100

## (2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 36 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

GATATGTCGA CGATGATGGA TATTTACGTA TGTTTG

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 48 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

GCATGGATCC ACCTCCTCCA GAATTTTCCT TTGTCACCAC ATACATAC

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 49 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CGCGTCCGGA GGAGGTGGAT CTGAACTTCT AGATCCATGT GGTTATATC

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 37 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

GCATGCGGCC GCCTATTCAA TTTCTCCTTG AGCAAAC

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 35 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

GATATGTCGA CAAGATGTTG ACGTTGCAGA CTTGG

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 41 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

GCATGGATCC ACCTCCTCCT TCAATTTCTC CTTGAGCAAA C

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 44 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

CGCGTCCGGA GGAGGTGGTA GCCAGAAAAA GGGGGCTCCT CATG

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 43 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GCATGCGGCC GCTAAGAATT TTCCTTTGTC ACCACATACA TAC

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 45 base pairs
- (B) TYPE: nucleic acid

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(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

GCATAGATCT GGGCTCAGAA TTTTCCTTG TCACCACATA CATAC

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 38 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

GCATAGATCT GGGCTCTTCA ATTTCTCCTT GAGCAAAC

CLAIMS

What is claimed is:

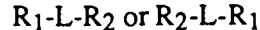
5

1. A receptor capable of binding oncostatin M and leukemia inhibitory factor, comprising gp130 covalently linked to LIF-R.

10 2. A receptor according to claim 1, wherein said receptor comprises a soluble gp130 polypeptide covalently linked to a soluble LIF-R polypeptide.

3. A receptor according to claim 1 wherein said receptor comprises gp130 covalently linked to LIF-R *via* a polypeptide linker.

15 4. A receptor according to claim 3, wherein said receptor is a recombinant fusion protein of the formula:



20 wherein R<sub>1</sub> represents gp130; R<sub>2</sub> represents LIF-R; and L represents a polypeptide linker.

25 5. A receptor according to claim 4 wherein the polypeptide linker comprises from 20 to 100 amino acids selected from the group consisting of glycine, asparagine, serine, threonine, and alanine.

6. A receptor according to claim 5 wherein the polypeptide linker comprises an amino acid sequence selected from the group consisting of :  
(Gly<sub>4</sub>-Ser-Gly<sub>5</sub>-Ser)<sub>2</sub> and (Gly<sub>4</sub>-Ser)<sub>n</sub>, wherein n is 4-12.

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7. An isolated DNA sequence encoding the receptor of claim 4.

8. A recombinant expression vector comprising the DNA sequence of claim 7.

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9. A host cell containing the expression vector of claim 8.

10. A receptor according to claim 3, comprising a first fusion polypeptide that comprises an antibody Fc region polypeptide attached to the C-terminus of gp130, and a second fusion polypeptide that comprises an antibody Fc region polypeptide attached to the C-terminus of LIF-R, wherein said first fusion polypeptide is linked to  
5 said second fusion polypeptide *via* disulfide bonds between the Fc region polypeptides.

11. A receptor according to claim 1, 4, or 10, wherein:

a) said gp130 is encoded by an isolated DNA selected from the group  
10 consisting of a first DNA sequence comprising nucleotides 244-2369 of SEQ ID NO:1, a second DNA sequence comprising nucleotides 310-2369 of SEQ ID NO:1, and a third DNA sequence that will hybridize to said second DNA sequence under moderately stringent conditions; and

b) said LIF-R is encoded by an isolated DNA selected from the group  
15 consisting of a first DNA sequence comprising nucleotides 179-3182 of SEQ ID NO:5, a second DNA sequence comprising nucleotides 311-3182 of SEQ ID NO:5, and a third DNA sequence that will hybridize to said second DNA sequence under moderately stringent conditions.

20

12. A receptor according to claim 10 wherein said gp130 is a soluble gp130 polypeptide and said LIF-R is a soluble LIF-R polypeptide.

13. A fusion protein comprising an antibody Fc region polypeptide attached  
25 to the C-terminus of a soluble gp130 polypeptide.

14. An isolated DNA sequence encoding a fusion protein according to claim  
13.

15. A fusion protein comprising an antibody Fc region polypeptide attached  
30 to the C-terminus of a soluble LIF-R polypeptide.

16. An isolated DNA sequence encoding a fusion protein according to claim  
15.

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17. A homodimeric receptor comprising two fusion proteins according to claim 15, linked *via* disulfide bonds between the Fc region polypeptides.

18. A process for preparing a receptor according to claim 4, comprising culturing a host cell transformed with an expression vector comprising a DNA sequence that encodes said fusion protein under conditions that promote expression of said fusion protein, and recovering said fusion protein.

19. A process for preparing a receptor according to claim 10, comprising culturing a host cell co-transfected with a first expression vector encoding said first fusion polypeptide and with a second expression vector encoding said second fusion polypeptide under conditions that promote expression of said first and second fusion polypeptides, and recovering said receptor.

20. A pharmaceutical composition for treating a disorder mediated by oncostatin M or LIF, comprising the receptor of claim 1, 4, 10, or 12, and a suitable diluent or carrier.

FIGURE 1

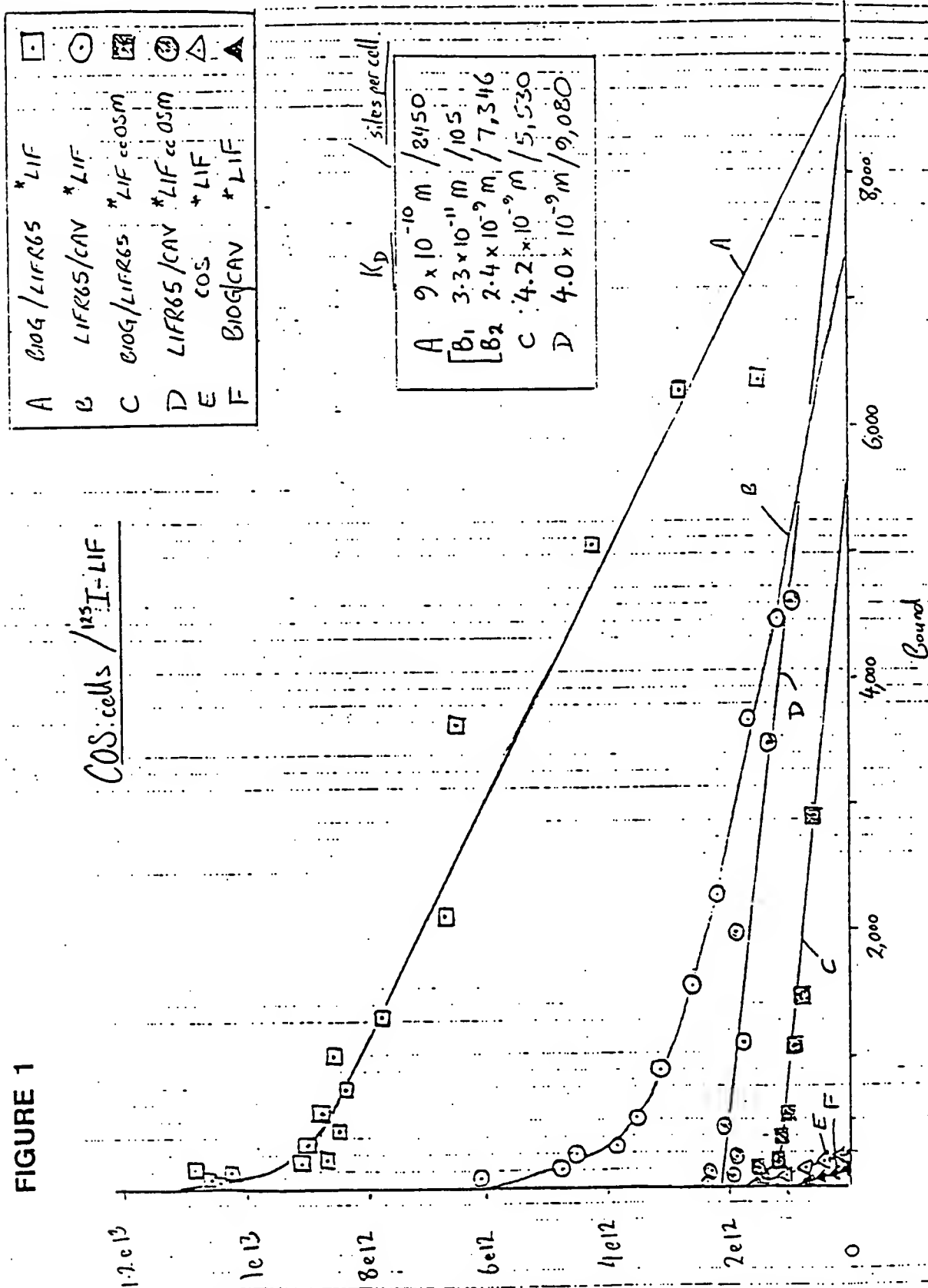


FIGURE 2

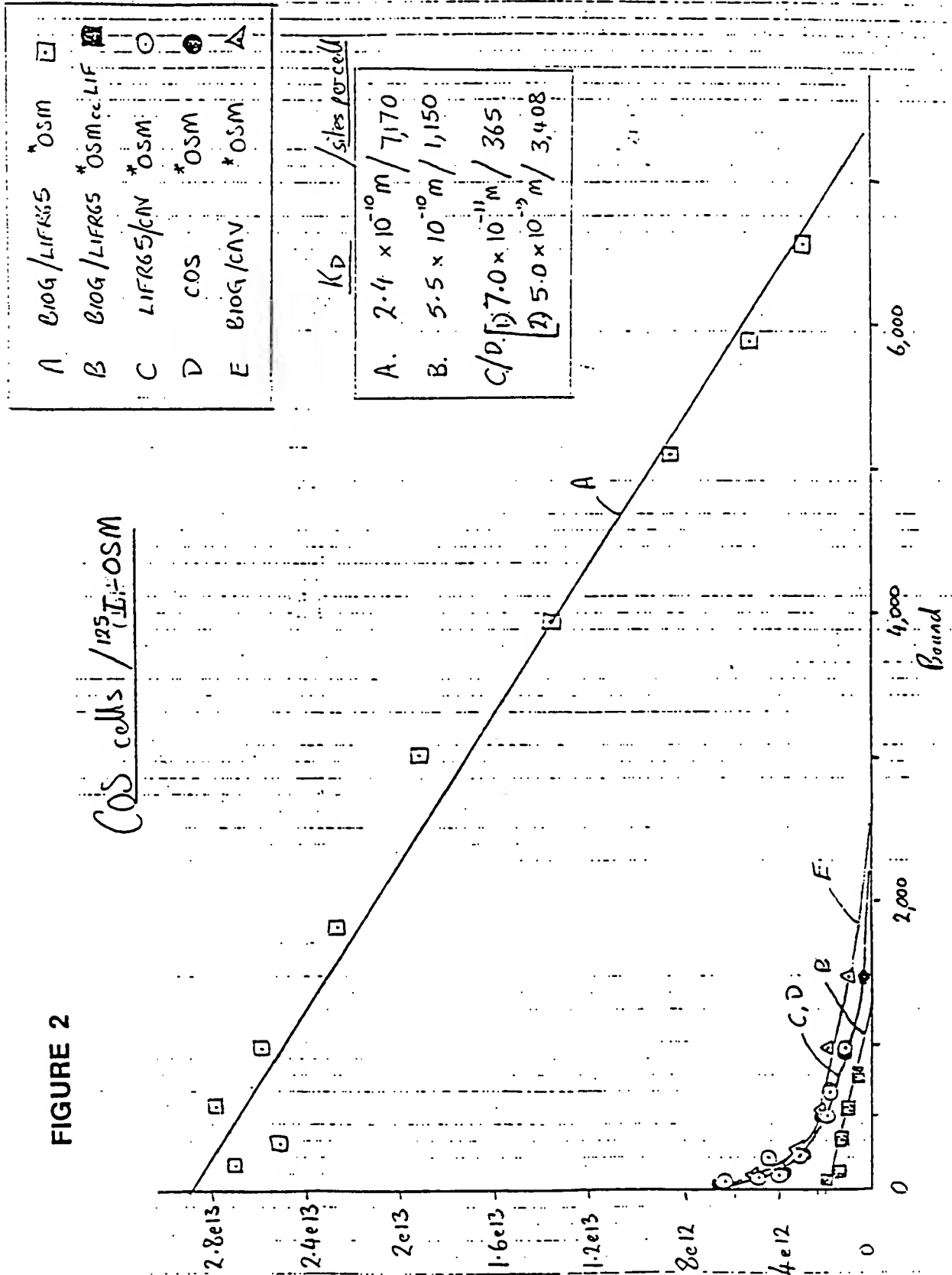


FIGURE 3

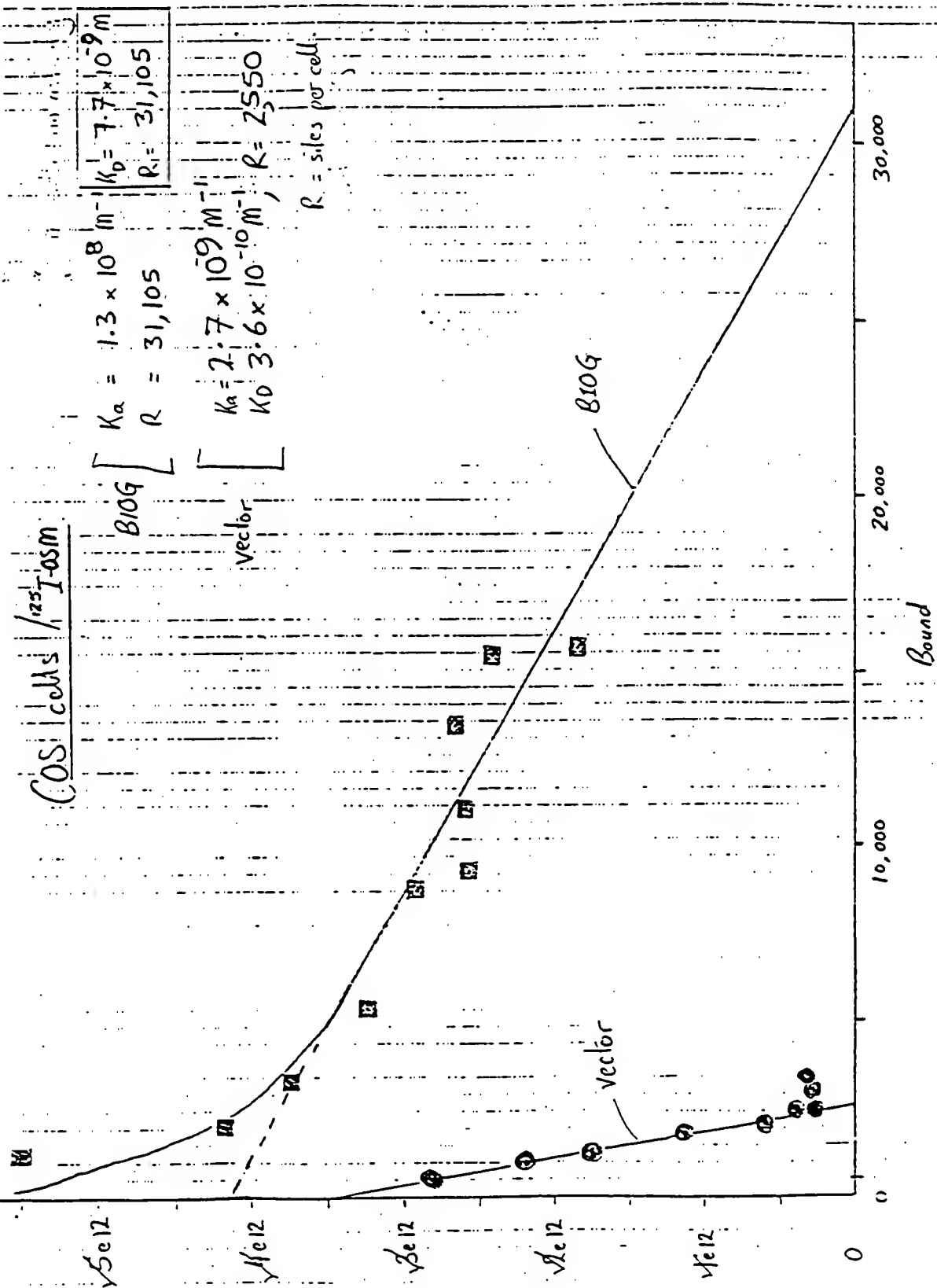
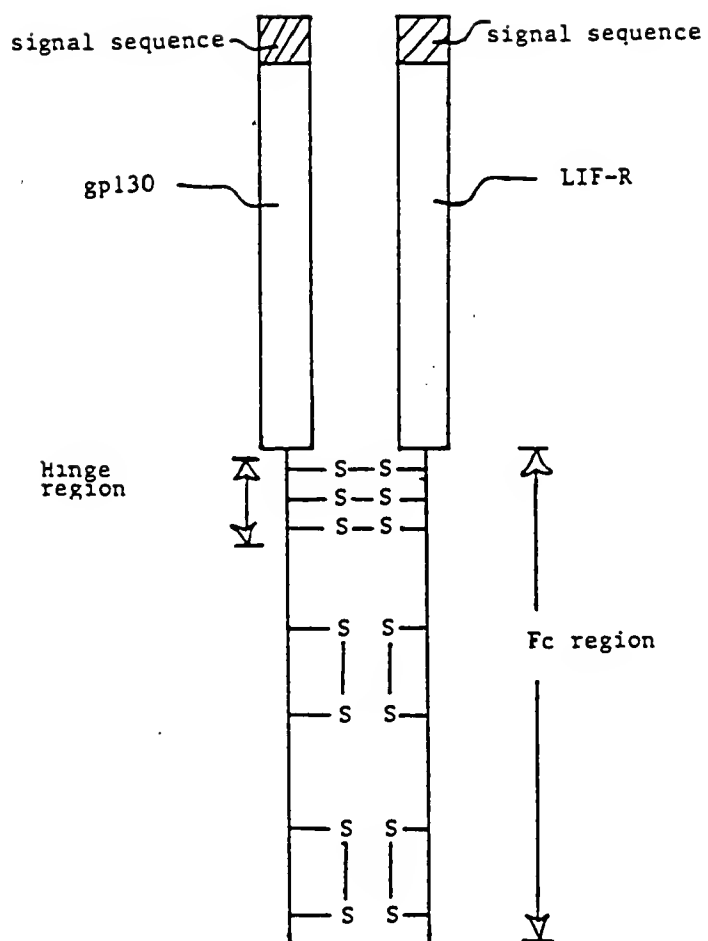


FIGURE 4



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FIGURE 5

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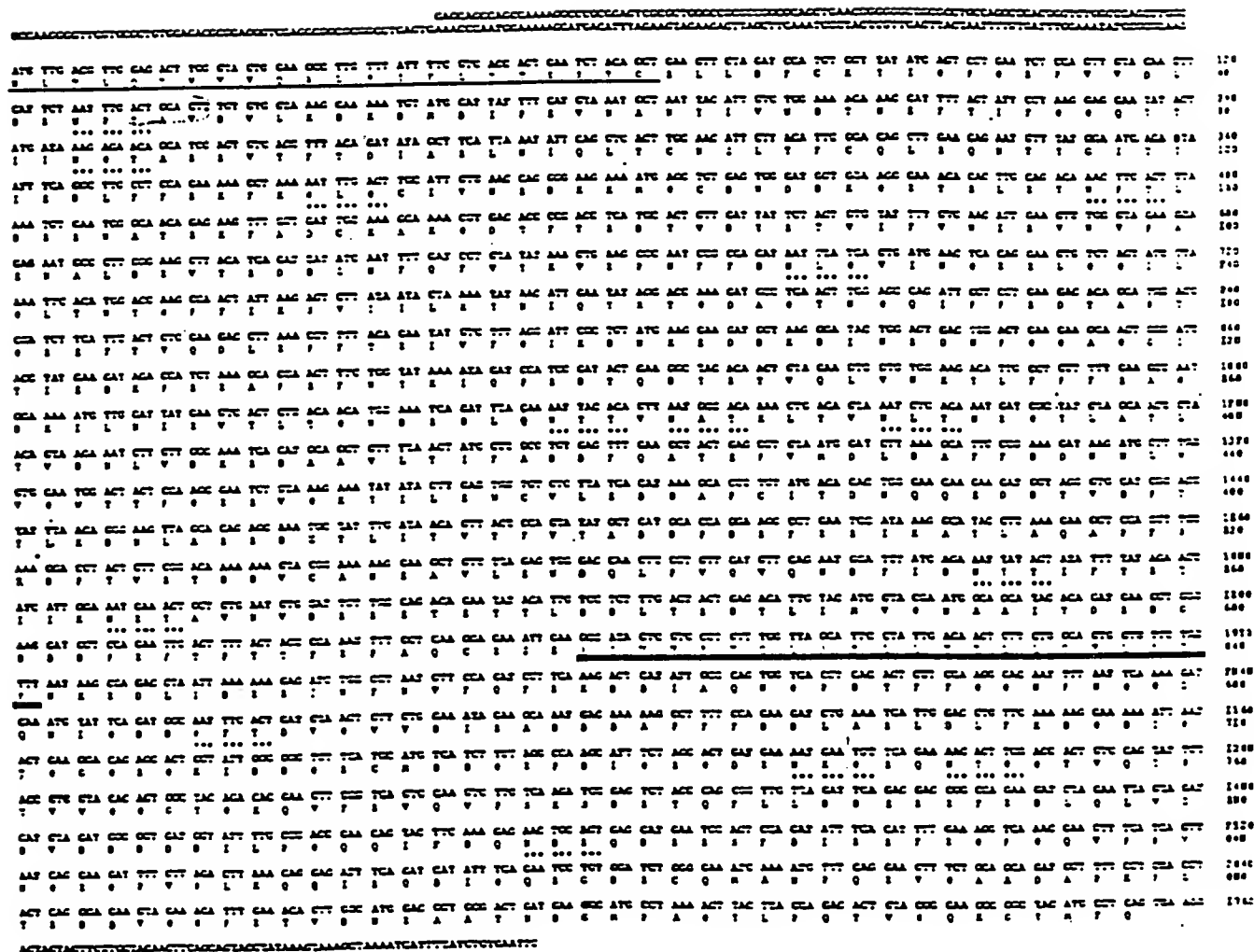
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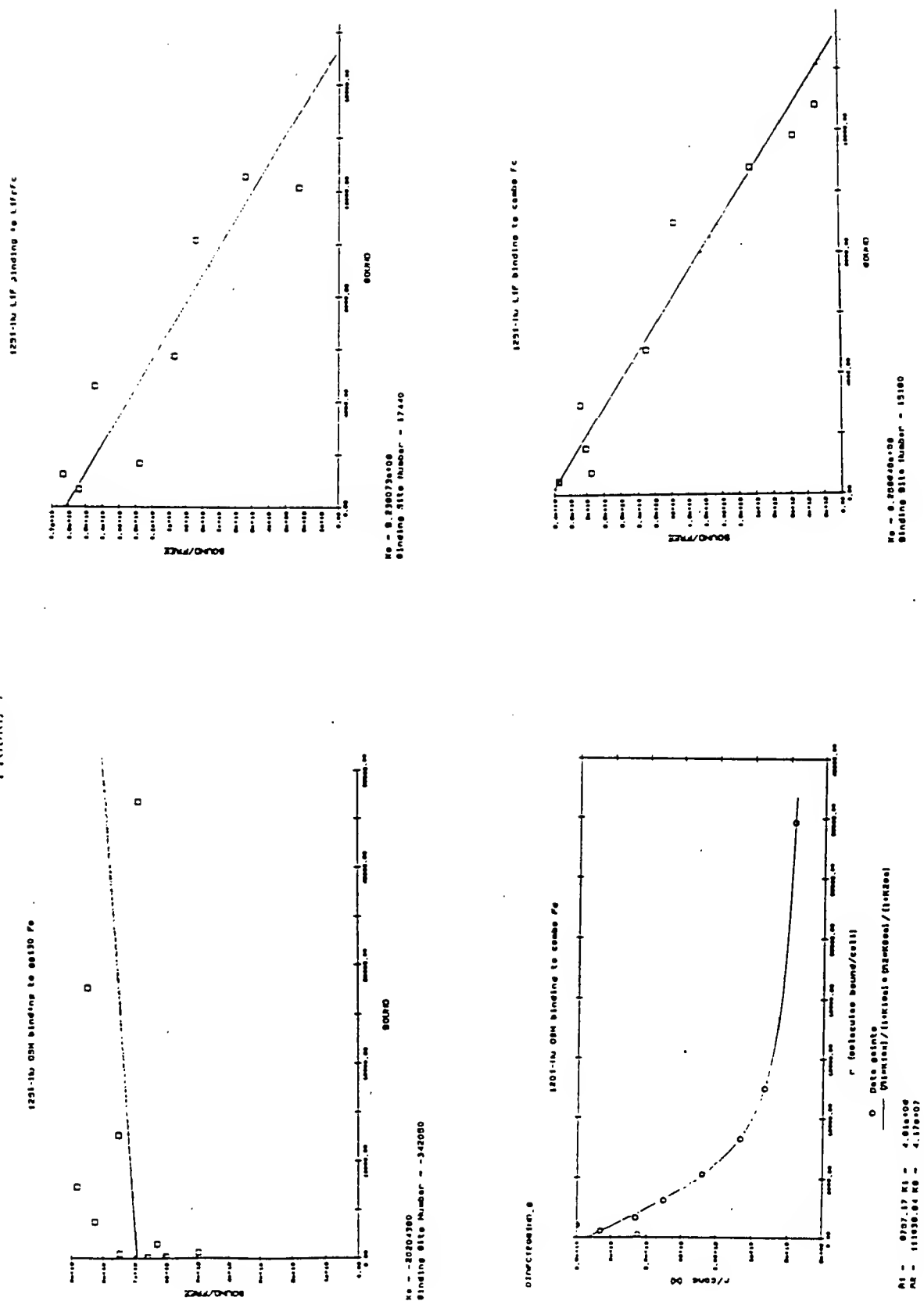
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FIGURE 6



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FIGURE 7



## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US92/10272**A. CLASSIFICATION OF SUBJECT MATTER**

IPC(5) : C07K 13/00; C12N 15/62

US CL : 435/60.7; 514/2; 530/350

According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/69.1, 69.7, 252.3, 320.1; 514/2; 530/350; 536/27

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, STN/MEDLINE

search terms: gp130, leukemia inhibitory factor, receptor#

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	EMBO Journal, Vol. 10, No. 10, issued October 1991, D. P. Gearing et al, "Leukemia inhibitory factor receptor is structurally related to the IL-6 signal transducer, gp130", pages 2839-2848, see entire document.	1-20
A	Cell, Vol. 63, issued 21 December 1990, M. Hibi, et al, "Molecular Cloning and Expression of an IL-6 Signal Transducer, gp130", pages 1149-1157, see entire document.	1-20
A,P	US, A, 5,155,027 (Sledziewski et al) 13 October 1992, column 5, line 38 to column 6, line 16.	1-20

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
*A* document defining the general state of the art which is not considered to be part of particular relevance	*X*	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
*E* earlier document published on or after the international filing date	*Y*	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
*L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*A*	document member of the same patent family
*O* document referring to an oral disclosure, use, exhibition or other means		
*P* document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

05 March 1993

Date of mailing of the international search report

10 MAR 1993

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